Genetically Modified Plants and "Non-Target" Organisms: Analysing the Functioning of the Agro-ecosystem

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Abstract

One of the main concerns for a possible adverse environmental effect due to the cultivation of genetically modified (GM) crops is the threat to the biodiversity in the receiving environments where such plants will be cultivated. In particular, animal biodiversity could be at risk if organisms that come in contact with GM plants and/or their products are harmed. In any ecosystem, including agro-ecosystems, hundreds of species are sustained in food webs, above and below ground, based on cultivated plants as the main primary producers. Therefore, numerous species at higher trophic levels can come in contact with plant metabolites either directly or indirectly. GM crops developed to control insect pests are purposefully hazardous to certain pest species considered to be the target of these new plant varieties (e.g. Cry toxin-expressing GM plants). All other organisms active in the agro-ecosystem and in adjacent habitats are not intended to be harmed by these GM plants and, as a consequence, can operationally be defined as non-target organisms (NTO). In this review the state of the art concerning possible effects of GM crop plants on NTOs is summarised according to the ecological role these organisms play in agro-ecosystems. This criterion is also suggested as a pathway for evaluating possible adverse effects on NTOs when GM crops are being considered for release into a new receiving environment or for the release of new GM events into well-characterised receiving environments. The idea underlining this proposal is that in heavily human-managed ecosystems, such as agro-ecosystems, the preservation of functional biodiversity is an important protection goal in ecological terms, but it is also paramount for the sustainability of agriculture.

Riassunto

Una delle principali preoccupazioni in merito ad un possibile effetto ambientale negativo dovuto alla coltivazione di piante geneticamente modificate (GM) è la minaccia per la biodiversità negli ambienti dove queste piante verranno coltivate. In particolare, la biodiversità animale potrebbe essere a rischio se gli organismi che vengono a contatto con le piante GM e/o con i loro prodotti sono danneggiate. In ogni ecosistema, inclusi gli agro-ecosistemi, centinaia di specie vengono sostenute all'interno di reti alimentari, nel sopra e nel sottosuolo, basate sulle piante coltivate quali principali produttori primari. Pertanto, numerose

specie ai livelli trofici superiori possono venire in contatto con i metaboliti di tali piante direttamente o indirettamente. Le piante GM resistenti agli insetti sono appositamente ottenute per indurre effetti negativi su alcune specie fitofaghe che sono quindi considerate come bersaglio di queste nuove varietà (es. piante GM esprimenti tossine Crv). Al contrario, queste piante non hanno lo scopo di interferire con nessuno degli altri organismi attivi nell'agro-ecosistema e negli habitat adiacenti che pertanto possono essere definiti in termini operativi guali organismi non-bersaglio (NTO). In questa review lo stato dell'arte nel campo degli studi degli effetti delle piante GM sugli NTO viene presentato considerando il diverso ruolo ecologico che tali organismi rivestono negli agro-ecosistemi. Questo stesso criterio viene anche suggerito come modalità operativa per valutare i possibili effetti sugli NTO in nuovi ambienti o in presenza di una nuova pianta GM. L'idea che sottende a questo approccio è che in ecosistemi fortemente antropizzati, quali gli agro-ecosistemi, il mantenimento della biodiversità funzionale rappresenta un importante obiettivo di protezione ambientale, ma è anche fondamentale nell'ottica della sostenibilità dell'agricoltura.

1. INTRODUCTION

Genetically modified plants (GMPs) currently in cultivation are principally designed to express characters of herbicide tolerance or insect resistance (IR). In the latter case, commercially available GMPs were produced to express resistance to some Lepidoptera or Coleoptera pest species (e.g. Ostrinia nubilalis Hubner, Helicoverpa zea (Boddie), Diabrotica virgifera virgifera LeConte, Leptinotarsa decemlineata Say, Chilo suppressalis Walker, etc.), mostly via the expression of modified cry genes originally derived from the soil bacterium Bacillus thuringiensis Berl. Indeed, pest species represent a small proportion of the organisms that may come into contact with IR crop plants in agro-ecosystems, therefore concerns have been expressed that commercial cultivations of GM crops could affect other possibly sensitive species.

The concept of "non-target" organisms (NTO) has become common in debating the biosafety of GMPs, the specific risk assessment of which is often required by law. For instance, the European Directive 2001/18/EC provides the legal background for NTO testing, requiring the assessment of possible changes in the interactions of GM plants with NTOs prior to their commercial release. The concept of NTO can only be defined in operational terms and in relation to traits newly expressed in plants, as there is no correspondence with any ecological role of these organisms in their ecosystems. In this article, NTOs will be considered as "all living organisms that are not meant to be affected by newly expressed compounds in GMPs, and that can be potentially exposed, directly or indirectly, to the GM plant and/or its products in the agro-ecosystem where GMPs will be released or in adjacent habitats". However, an additional case is represented by organisms indirectly affected via changes in the cultivation, management and harvesting techniques as a result of the introduced trait in GMPs, for example changes to herbicide application regimes when cultivating herbicide tolerance plants. A few examples of such cases will also be highlighted in this review. For GM plants expressing traits that do not confer resistance to pests, all organisms potentially exposed are therefore considered as non-target. Most of the chosen examples in this paper will refer to arthropods; relevant implications regarding other organisms will be highlighted and the appropriate references will help readers to further reflect on these issues.

Agriculture depends on several ecosystem functions that are essential to soil fertility and agricultural productivity (e.g. microbial decomposition and nutrient cycling, crop pollination by animals, biological control of pests). Each of these ecosystem functions are mediated by several guilds of animal species. Therefore, in any given cropping system, many hundreds of arthropod species, thousands of microbial species, and scores of ecosystem functions can be found (Curtis *et al.*, 2002). It is therefore impossible to estimate the impacts of any new cultivation technique on all potentially exposed species.

To date, the environmental release of GMPs takes place on commercial farms in rural areas. Therefore the concept of the agro-ecosystem (i.e. a specialised, human-managed ecosystem designed for the production of agricultural goods) and its mode of functioning can represent a useful logical criterion to guide the analysis of the large trophic webs that exist therein and where the potential exposure of NTOs to newly produced compounds in GMPs can occur.

Food webs in agro-ecosystems are typically simplified compared to natural habitats, due to the major impact of human activities, the short time span plants remain in the field and the usual uniformity of cultivated plants. Even so, rather complex multi-trophic relationships are established between primary producers and consumers, at least up to the fourth trophic level (e.g. Verkerk, 2004).

Many reviews are already available on the subject of possible effects of GMPs and NTOs seen from different view points. Among the most recent, O'Callaghan *et al.* (2005) gave an overview of the available knowledge concerning the effects of IR GMPs on plant- and soil-dwelling arthropods. Lövei & Arpaia (2005)

concentrated on natural enemies, analysing individual response classes observed in laboratory studies and highlighting several existing knowledge gaps. Romeis *et al.* (2006) summarised the overall effects of Cry toxins on natural enemies using conclusions from laboratory and field studies, as opposed to effects due to chemical pesticides. An updated review by the same group (Romeis *et al.*, 2009) highlighted the lack of direct effects of this group of toxins on many non-target above-ground arthropods and advise that future GM crops should be assessed on a case-by-case basis. Malone & Burgess (2009) examined the existing literature on Hymenoptera pollinators and GMPs, of which some of their results will be commented upon later (Section 3.2.).

The use of meta-analysis to summarise available data has been recently adopted in the reviews of field studies in Bt-expressing maize and cotton (Marvier et al., 2007), functional groups of non-target arthropods (Wolfenbarger et al., 2008), specific effects on honey bees (Duan et al., 2008), and the occurrence of direct and/or indirect effects (Naranjo, 2009). Lövei et al. (2009) concentrated their metaanalysis on laboratory studies and natural enemies, performing calculations based on single endpoints measurements and using effect size classes instead of an average effect size. The conclusions of these meta-analysis studies are discussed later (Section 3.1.). Soil communities, including microorganisms, have also been reviewed in recent papers. Widmer (2007) and Filion (2008) concentrated specifically on microbial communities while soil associated meso- and macrofauna were also considered by lcoz and Stotzky (2008). Together, these analyses concluded that GMPs will likely induce changes in these communities, but these are expected to be within the range of natural variation (Section 3.3.6).

The present review aims to summarise the specific information available in the literature regarding NTOs and GMPs, particularly those which express insect resistance characters. The review will not try to give opinions about the environmental safety of specific transformation events, but will analyse information and information gaps concerning the main functional guilds active in agroecosystems and adjacent habitats. The main biological mechanisms regulating food webs will be reiterated and used to evaluate the possible exposure routes to hazardous compounds that may be expressed in GMPs. The available scientific knowledge will be revisited with the specific aim of "learning lessons" and trying to help researchers, risk assessors and regulators in different parts of the world, including those where specific information is limited, to collect the most relevant data necessary for an effective and scientifically-sound environmental risk assessment.

2. HOW CAN NON-TARGET ORGANISMS BE AFFECTED BY GENETICALLY MODIFIED PLANTS?

The concept of non-target organisms may include a large number of different species. Therefore there are different reasons why the impacts of GMPs on NTOs in agricultural fields and nearby habitats are commonly perceived as possibly serious threats. These possible negative effects can be categorised, according to the functions or values that can be affected, as follows: ecological effects, effects on agriculture, and effects on other anthropocentric values.

Ecological effects relate to threats to biodiversity and ecological functioning in the ecosystems. The protection of biodiversity is deemed important for evolution and for maintaining life sustaining systems of the biosphere, and it is a common concern of humankind (Secretariat of the Convention on Biological Diversity, 1992). Generally, environmental protection goals aim to protect biodiversity through the conservation of natural habitats, wild fauna and flora (e.g Directive 92/43/EEC). Since GMPs will be released in cultivated areas, a possible deterioration of cropping system needs to be considered. The effects on farming activity are related to the biodiversity effects, particularly when "ecosystem services" (Daily, 1997) provided by different guilds can possibly be impaired (e.g. should animal pollination be threatened by the expression of new proteins in crop plants, or natural biological control of pests reduced because toxins accumulate along trophic chains, etc.).

The same biological events can induce, for instance, consequences at different levels. For example, gene flow is a common event in nature and constitutes the base of evolution for every environment, therefore it cannot be considered a negative event *per se*. To evaluate its ecological consequences, it is important to consider the possibility of successful fertile hybrid formation and the successive introgression of a new trait into the population. This latter event is normally foreseeable as long as it confers a selective advantage to the progeny in the specific receiving environment. A possible negative ecological effect, as a consequence of gene flow, could be considered an increase in the invasiveness of a wild plant acquiring a GM trait as a consequence of natural crossing. The corresponding effect on farming activities could be the possible development of an herbicide resistant weed which could affect crop yield due to its newly acquired trait (Lu, 2008). Finally, other anthropocentric values can be threatened, such as the survival of rare, endangered or charismatic species (e.g. *Danaus plexippus* L., the Monarch butterfly in the USA) or the normal development of economic activities, such as apiculture.

The scientific approach to risk assessment applied in many different fields involves the consideration of two broad categories; hazard characterisation and exposure evaluation. An environmental risk due to release of GMPs indicates the probability of an adverse effect occurring due to the exposure of NTOs to the GM plant and/ or its products. In the following chapters, hazard characterisation and evaluation of exposure will be analysed in relation to agro-ecosystems based on the most current scientific literature.

3. A "FUNCTIONAL" APPROACH

As outlined above, the large number of species present in any agro-ecosystem renders impossible any detailed study including all of them. Therefore, it is necessary to make a choice of assessment endpoints that could be considered representative for the specific receiving environment.

Many possible criteria have been proposed, which usually enable the selection of a handful of species based on characteristics such as their abundance in the specific environment, the susceptibility to known stressors, the practicability of conducting laboratory tests, the charismatic value of the species, etc. (e.g. Cowgill and Atkinsons, 2003; Andow and Hilbeck, 2004; Birch *et al.*, 2004; Prasifka *et al.*, 2008; Romeis *et al.*, 2008; Todd *et al.*, 2008). Therefore, it is not surprising that the available body of data does not evenly represent the various animal taxa linked to the agro-ecosystem, and that not every specific guild was studied to the same extent.

Ecosystem services denote ecological processes that benefit humankind (Daily, 1997). Several types of ecosystem services ensure agricultural productivity, including soil formation, decomposition of plant residues, pollination, and natural pest control, to name a few. Regardless of GMO introductions, several of these services are already considered to be under pressure and their ability to continue at desired rates is at risk (MA, 2005). The present narrative review of the scientific literature will revise the available knowledge regarding functional groups (i.e. all clusters of biota providing the same ecological function) of organisms active in agro-ecosystems and adjacent habitats (Table 1.) and link each of these groups to ecosystem services and to the main anthropocentric values. The idea behind

this choice is that it can be assumed that a change in biodiversity structure may result in a change in function; therefore, preserving functional biodiversity may guarantee the quality of agro-ecosystems. In fact, increasing biodiversity is known to enhance agro-ecosystem resilience and stability in the presence of redundant species and, more importantly, to increase ecosystem functioning in terms of processes or magnitude of processes in species-poor agro-ecosystems in the short term (Moonen and Bàrberi, 2008). Moreover, not every species or process is equally important for the functioning of the agro-ecosystem and therefore a preliminary problem formulation may successfully use this approach to prioritise which species or functions to be analysed.

The first areas that will be potentially affected by the deployment of GMPs are the cultivated fields and their surroundings. From an applied perspective, we might then essentially concentrate on functions that are important in such environments. Moreover in habitats adjacent to farms, floral and faunal biodiversity may also represent important anthropocentric values to be protected. In the agroecosystem, crop plants, weeds and volunteer plants in the field and field margins are included. Adjacent habitats are those where plant parts can be present on a regular basis (e.g. via pollen drift or cultivation residues). This has obviously some limitations, but will likely include the most relevant mechanisms possibly leading to impacts. In relation to agro-ecosystems, the main ways in which a non-target species can be exposed to GMPs or their products are via air, plant and soil. Table 2 summarises the relative exposure mechanisms linked to these elements and the non-target groups possibly involved.

Table 1. Examples of functional group of species and ecosystem services provided	Ĺ
to human activities	

		Examples of
Functional group	Examples	ecosystem services or
		anthropocentric values
Herbivores	 Sap/cell feeders: e.g. Acarina Tetranychidae, Homoptera: Aphidioidea, Aleurodidae, Cicadellidae; Thysanoptera: Thripidae, etc. Leaf feeders: e.g. Coleoptera: Chrysomelidae, Lepidoptera larvae, etc. 	Secondary pests, quarantine pest species, species of conservation concern, species of charismatic value
Predators	Thysanoptera (e.g. Aeolothripidae); Heteroptera (e.g. Anthocoridae, Miridae, Nabidae); Neuroptera (e.g., Chrysopidae, Hemerobidae); Coleoptera (e.g. Coccinellidae, Carabidae, Staphilinidae); Diptera (e.g., Syrphidae, Cecidomyiidae); Araneae; Acarina (e.g., Phytoseiidae)	Natural control of arthropod pests
Parasitoids	Diptera (e.g. Tachinidae); Hymenoptera Parasitica	Natural control of arthropod pests
Pollinators, pollen feeders	Diptera: Syrphidae; Hymenoptera: Apidae	Crop plant pollination, honey production
Decomposers	Nematoda, Collembola, Acarina, Haplotaxida: Lumbricidae, Isopoda	Soil fertility

Exposure channel	Mechanism	Non target groups	Ecosystem services possibly affected
AIR	Pollen flow, seed dispersal	 Sexually compatible plants (gene flow); Herbivores ingesting pollen in field margins and adjacent habitats; Spermophagous species in adjacent habitats 	• Pollination
PLANT	Trophic chain effects	 Primary consumers (herbivores) in the field; Higher order consumers (carnivores) in the field and in adjacent habitats. These groups are not limited to arthropods, but may include e.g. rodents, birds, mammals, etc. 	PollinationNatural pest control
SOIL	Horizontal gene transfer, release of plant products	Microorganisms, meso- and macro-fauna	Soil fertilityNatural pest control

 Table 2. Possible exposure mechanisms and their consequences for non-target organisms

3.1. Natural Pest Control

It is estimated that about 95 % of the potential pest arthropod species are maintained below damaging population levels by natural pest control provided by predators and parasitoids (DeBack and Rosen, 1991). This is a typical regulating service provided by numerous animal guilds that prevent most of the herbivores living in agro-ecosystems in becoming pests. The maintenance of this ecosystem service is estimated to have a minimum value of 400 billion USA dollars per year (Costanza *et al.*, 1997). Moreover, natural pest control constitutes a "baseline" value over which any integrated pest management (IPM) programme is built,

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since the population dynamics of herbivores and their natural enemies are routinely monitored before any pest management action is considered. It is clear then that preserving this ecosystem service is paramount in order to guarantee the sustainability of agriculture, and to limiting the need for chemical pesticides.

Plants expressing IR characters could introduce novel metabolites into existing food web (e.g. Cry toxins, proteinase inhibitors, lectins), therefore higher order consumers may possibly be adversely affected. The most widely cultivated IR GM crops contain modified versions of genes derived from the soil bacterium B. thuringiensis (Bt). There is also case of a commercial GM plant expressing a serine protease inhibitor in combination with a Cry toxin in cultivation in China (Malone et al., 2008). These genes enable the production of crystal proteins (Cry) which are selectively toxic to various insect orders. Many different strains of B. thuringiensis were isolated from soil samples, plant surfaces, dead insects and stored grains from all over the world. The different strains show a wide range of specificity against different insect orders (Lepidoptera, Diptera, Coleoptera, Hymenoptera, Homoptera, Phthiraptera/Mallophaga), Acarina, Nematoda and other invertebrates (http://www.glfc.forestry.ca/bacillus/BtSearch.cfm). The assumed selectivity of Cry toxins is therefore an important characteristic that should limit the negative effects of GM plants to target species. In some cases though, the range of sensitivity to Cry toxins is not completely known.

3.1.1. Predators

Arthropod predators are very active in the natural biological control in agroecosystems; in about 75% of cases generalist predators, either single species or species assemblages, reduce pest numbers significantly (Symondson et al., 2002). To date, 41 predator species (Table 3) have been considered in laboratory studies in relation to IR GMPs. A large part of the available information though, relates primarily to three predatory species, Chrysoperla carnea Steph. (Neuroptera: Chrysopidae), Propylea japonica Thunberg (Coleoptera: Coccinellidae), and Coleomegilla maculata DeGeer (Coleoptera: Coccinellidae) (Lövei et al., 2009). The limited species spectrum is not the only existing shortcoming in the available data, but also sample size, statistical power and duration of certain laboratory toxicity tests are limiting factors in several papers (Lövei and Arpaia, 2005). By analysing these laboratory studies, Lövei et al. (2009) indicated that there are fewer neutral responses, and more positive and negative effects than expected under the assumption of normal distribution of the responses. This would generally translate into the indication of non-neutral effects for both Cry toxins and proteinase inhibitors, the latter giving more significantly negative or positive

effects. Cry3A/Bb generally induced fewer effects in either direction compared to other toxin classes.

Naranjo (2009) also considered laboratory studies on predators but specifically distinguished between direct or prey-mediated effects. When direct-feeding experiments alone were considered, predators showed a significant reduction in their developmental rate when exposed to *Bt* proteins compared with non-*Bt* controls. Conversely, *Bt* toxins had no affect on the survival or reproduction of predators. When experiments only addressing prey-mediated effects were considered, predators showed slightly lower survivorship when provided with low-quality prey exposed to *Bt* toxins, and slightly faster developmental rates when provided with high-quality (i.e. not susceptible to *Bt*) prey exposed to *Bt* toxins. All other predator life history characteristics were unaffected by *Bt* toxins regardless of prey quality.

Whenever a possible hazard is identified in laboratory studies, it is generally advised to further investigate possible environmental impacts in more realistic conditions under more natural exposure conditions in semi-field or field studies. Negative and positive aspects of field studies, as opposed to laboratory tests, are briefly outlined in Section 5. Specific field studies have been conducted on several GM crops. Marvier *et al.* (2007) summarised them by undertaking a meta-analysis of data collected from various field studies of *Bt*-expressing maize and cotton in different parts of the world. They showed that the combined abundance of all non-target invertebrates was significantly lower in *Bt* crops if compared with non-*Bt* crops that had been treated with insecticides.

Wolfenbarger *et al.* (2008) also analysed field studies using arthropod abundance as the measurement endpoint. In this meta-analysis, the authors also included studies with Cry3-expressing GM potatoes, moreover their results were obtained after pooling experimental data according to the functional guilds of the NTO species monitored. The authors showed that in cotton, there were significantly fewer predators in *Bt* crop fields compared to un-sprayed, control (non-*Bt*) fields. This result was not related though to the feeding style within this functional group, but was largely driven by the lower abundance of Nabidae and Coccinellidae found in Bt crop fields. Higher numbers of the generalist predator, *C. maculata*, were associated with *Bt* maize but numbers of other common predatory genera were similar in *Bt*

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and non-*Bt* maize. Finally, in the case of the GM potatoes, there were more predators and herbivores in Bt crop fields than in un-sprayed control fields. The analysis of this latter crop though, included only two field studies and significant heterogeneity existed in both of these functional groups.

Cloutier et al. (2008) were specifically interested in Bt potato resistant to the Colorado potato beetle and used published studies to perform a specific metaanalysis. The authors found in about 20 % of cases an increased abundance of generalist predators, and suggested that this result could be related to the rather common (42% of cases) higher abundance of sucking herbivores detected on Bt potato. The exposure of predators to Cry toxins expressed in GM plants has been demonstrated experimentally. Many studies are now available which confirm that predators can ingest newly-expressed proteins in GMPs by direct feeding on Btexpressing plant material (e.g. pollen), and indirectly through the consumption of herbivore prey. Harwood et al. (2005) studied the exposure of several groups of non-target organisms to the Cry1Ab toxin from Bt11 maize and reported its observed levels in non-target herbivores and their natural enemies under field conditions. Significant quantities of the Cry1Ab toxin were detected in organisms at higher trophic levels. Obrist et al. (2006a) showed that the Cry1Ab toxin from Bt176 maize in some cases accumulated in the food chain at concentrations higher than those in the maize leaves. The Cry1Ab toxin was detected in certain predators (such as Orius spp., Chrysoperla spp. and Stethorus sp.), whilst its presence was negligible in others (e.g., Hemerobiids, Nabis sp., Hippodamia sp., Demetrias sp.). This difference might be the result of the different feeding habits of prev species which can indest variable amounts of toxins. In another study, Obrist et al. (2006b) confirmed toxin uptake by larvae of green lacewing, C. carnea, via its herbivore prey, Tetranychus urticae Koch and Spodoptera littoralis Boisduval; the biological activity of the Cry1Ab toxin was maintained after ingestion by both herbivore species. Harwood et al. (2007) showed the presence of the Cry1Ab toxin in gut samples of certain predatory coccinellids (e.g., C. maculata, Harmonia axyridis –Pallas-, Cycloneda munda –Say– Coccinella septempunctata L.). Álvarez-Alfageme et al. (2008) detected Cry1Ab toxin in the coccinellid species Stethorus punctillum Weise collected from MON810 maize fields. Generalist predators may in some cases show more complex feeding habits including feeding on plant parts, on numerous herbivore prey species and also intra-guild predation; in some cases with clear preferences among available food sources. These habits make the possible exposure to plant metabolites guite variable and difficult to predict.

3.1.1.1. Details about some taxonomic groups

Lacewings (Neuroptera: Chrysopidae)

As indicated above, the number of studies conducted on the green lacewing C. carnea has greatly outnumbered that of any other predatory species. More than ten years ago a series of publications (Hilbeck et al., 1998a; Hilbeck et al., 1998b; Hilbeck et al., 1999) reported significantly prolonged larval development and increased mortality when immature C. carnea were fed lepidopteran larvae reared on Cry1Ab expressing maize in laboratory. These findings have triggered numerous other studies on the same species. No acute adverse effects were reported when C. carnea larvae were fed non-susceptible T. urticae containing large amounts of biologically active Cry1Ab toxin (Dutton et al., 2002). Romeis et al. (2004) indicated that possible indirect effects may occur to this species due to poor prey quality when lepidopteran larvae are used as prey. Rodrigo-Simón et al. (2006) reported that Cry1Ab toxin does not specifically bind in vitro to brush border membrane vesicles from the midgut of C. carnea larvae, which is considered as a prerequisite for toxicity. Andow et al. (2006a) summarised the body of evidence available regarding this predator species, stating that studies agree that no acute toxicity of Cry1Ab to the larvae was demonstrated, when administered either in an artificial diet or in water. However, the laboratory experiments completed on this species do not completely exclude the possibility of chronic effects, as suggested by Hilbeck et al. (1998b).

Under field conditions, the continuous exposure of *C. carnea* to diets exclusively based on lepidopteran larvae is considered unlikely, since a variety of prey species is normally available (Dutton *et al.*, 2003). No differences in the abundance of lacewings in *Bt*-expressing and near isogenic control plots were observed in several field studies (e.g. Pilcher *et al.*, 1997; Wold et al., 2001; Bourguet *et al.*, 2002). In the meta-analysis conducted by Wolfenbarger *et al.* (2008), the weighted mean effect size based on field abundance for this species was essentially zero in both cotton and maize studies where no insecticides were used.

Ladybirds (Coleoptera: Coccinellidae)

Dhillon and Sharma (2009) studied the effects of Cry1Ab and Cry1Ac proteins on the predatory coccinellid *Cheilomenes sexmaculatus* –L.- under direct and indirect exposure conditions. Direct exposure of *C. sexmaculatus* larvae to *Bt* proteins at high concentrations resulted in reduced larval and adult emergence, as compared to controls. However, there were no adverse effects of the Cry toxins when the larvae were reared on *Aphis craccivora* Koch previously fed different concentrations of Cry1Ab or Cry1Ac in an artificial diet. The tritrophic experiments of Álvarez-Alfageme *et al.* (2008) demonstrated that when *S. punctillum* were fed *T. urticae* reared on transgenic *Bt*-expressing maize (events Bt176 and MON810), there was no effect on the survival of neonate ladybird larvae through to adulthood. Larval development was similar between treatments even though the fourth instar lasted longer for *Bt*-exposed larvae. Adult *S. punctillum* emerging from the feeding trials were further monitored for a measure of their relative fecundity. The results showed that the *Bt* maize of either event had no significant effect on the mean cumulative ladybird fecundity.

In other papers (e.g. Kalushkow & Hodek, 2005; Ahmad et al., 2006) no significantly negative effects on survival, growth and development of coccinellids were detected using two different Cry3 type of toxins. Zhang et al. (2006a) reported a decrease in body mass when Propylaea japonica Thunberg young larvae were fed for 72 h with 24 h old Spodoptera litura Fabricius larvae reared on Bt cotton cultivars GK-12 and NuCOTN 33B (expressing the fused Cry1Ab/Ac toxin and Cry1Ac toxin, respectively), compared to those fed with control-reared S. litura. Significantly fewer P. japonica larvae molted into second-instar when fed with S. litura reared on one of the Bt cotton lines used, compared to those fed prey from control plants. In laboratory feeding experiments using transgenic Bt cotton plants expressing the fused Cry1Ab/Ac toxin or Cry1Ac toxin, Zhang et al. (2006b) found that there were no distinct differences in pre-imaginal mortality between predators reared on aphids feeding on Bt cotton or control plants. The pre-imaginal stages of the ladybird beetles also developed faster when reared on prey fed Bt cotton cultivars than those fed control prey. However, there was a trend of more adult malformations when the predator was fed with prev from one of two Bt cotton cultivars than on control prey. Conversely, there were no significant differences in the pre-ovipositing period or in fecundity. Ladybird beetles preving on Bt-reared aphids matured faster and mated more frequently than those fed on aphids which were not exposed to Bt plants. When detailed analyses are performed, it is not always observed that all the measurement endpoints unequivocally indicate either the lack or presence of an effect. For instance, Bai et al. (2005) quantified and compared 18 predator response parameters of the effects of transgenic cry1Ab rice pollen on the fitness of the coccinellid P. japonica on two different Bt varieties Among the considered response parameters, two developmental parameters were significantly negative with respect to the control, whilst two other parameters (mortality and one developmental) were significantly positive. All the other data indicated a neutral effect of the diet.

Cry3 toxins are engineered in crop plants to confer resistance to Coleoptera,

therefore coccinellids that belong to the same order could theoretically be affected. Lövei et al. (2009) summarised all the published results from laboratory studies on C. maculata DeGeer, one of the two most commonly studied ladybird species in biosafety research with GMOs, and discovered that none of the 101 biological parameters investigated gave significantly negative results. A recent paper by Schmidt et al. (2009) reported on laboratory toxicity tests with microbially-produced trypsin-activated Cry1Ab or Cry3Bb proteins fed to different larval stages (L1-L4) of the coccinellid Adalia bipunctata L. Bt proteins were sprayed on Ephestia sp. eggs, which were then offered as food in a nochoice test to first instars of the study ladybirds. The authors report that A. bipunctata larvae fed lepidopteran-active Cry1Ab toxin exhibited significantly higher mortality levels than the control group. However, in experiments with the coleopteran-active Cry3Bb toxin, a higher concentration of the toxin (compared to Cry1Ab) was necessary to induce significant mortality levels. No differences were detected with respect to both development time of larvae and body mass of newly-emerged adults. The authors suggest that the increased mortality of larvae in the toxin feeding trials was caused directly by the activated Bt toxins and raise questions regarding their postulated specificity and mode of action in A. bipunctata. The higher toxicity on Coleoptera of a Lepidoptera-specific Cry1Ab in comparison to Coleoptera-specific Cry3Bb is a new outcome that needs to be confirmed based on more quantitative data of food intake and actual toxin concentrations in the diet.

Coccinellids have been the subject of several field studies with GMPs. No adverse effects of various Bt maize events were detected on a range of coccinellid species (e.g., Pilcher et al., 1997; Jasinski et al., 2003; de la Poza et al., 2005; Lundgren and Wiedenmann, 2005; Eckert et al., 2006). Wold et al. (2001) did not find adverse effects on A. bipunctata in field studies, but reported a lower abundance of C. maculata in GM sweet corn expressing Cry1Ab toxin in experimental fields. An important consideration in terms of environmental risk assessment is that it is unlikely for many coccinellid species to be exposed to biologically relevant amounts of Cry toxins from GMPs. This is primarily because aphids, as one of the preferred preys for ladybirds, are known to contain no (e.g. Head et al., 2001; Raps et al., 2001) or limited amounts (Zhang et al., 2006b; Burgio et al., 2007) of Cry toxins. Moreover, the Cry1Ab toxin content in commercialised GM maize pollen (which represents the most likely source for possible toxin ingestion for coccinellids) is usually limited in the most common events. For instance, Cry1Ab toxin in MON810 maize pollen ranges between 1-97 g/g fresh weight (Nguyen and Jehle, 2007). The exposure route used by Schmidt et al. (2009) may constitute

a useful model for laboratory studies, but any significant exposure through eggfeeding in the field can be considered very unlikely in field conditions.

Ground beetles (Coleoptera: Carabidae)

Because of their complex food webs and ground-dwelling habits, carabids may be exposed to transgenic Bt-endotoxins through multiple pathways, including root exudates (Saxena et al., 2002), soil-dwelling prey, ingestion of plant material, and pollen feeding. A relatively large number of ground beetle species have been subjected to specific studies with GMPs (Table 3). Peterson et al. (2009) studied the uptake of Cry toxins in carabids using several GMPs, including stacked events. The authors detected different amounts of toxins across predator species and attributed these differences to changes in the non-target food web as well as possibly to differential rates of Bt-endotoxin decay in the events studied. Álvarez-Alfageme et al. (2009) revealed that there was a great decline in the detection of Cry1Ab toxin through the trophic chain during tritrophic laboratory studies with Poecilus cupreus L. Meissle et al. (2005) reported a significant increase in the mortality of the generalist carabid predator P. cupreus L. fed S. littoralis larvae raised on Bt maize (event MON810). By speculating on the lack of specific receptors in *P. cupreus*, the authors assume the reported effects to be due to the nutritional guality of the prey and not to the direct toxicity of the Cry1Ab toxin, though they could not exclude direct toxic effects. The tritrophic experiments of Álvarez-Alfageme et al. (2009) also used P. cupreus larvae fed with S. littoralis caterpillars themselves previously fed either Bt176 or control maize. The results demonstrated that indirect exposure to Crv1Ab toxin had no effect on the developmental time of both larvae and pupae of *P. cupreus*. Similarly, no differences were observed on mortality and adult fresh weight. Riddick et al. (1998) reported a reduced abundance of Lebia grandis Hentz., a predator of the target pest L. decemlineata, in experimental fields with Coleoptera-resistant Cry3A-expressing potato. De la Poza et al. (2005) noted that the abundance of Carabidae varied from year to year and between locations during a three-year monitoring programme in Spain, but no clear relationships with Bt maize was discerned

Ground beetles were also one of the groups of non-target insects considered in the British farm-scale trials with genetically modified herbicide tolerant (GMHT) crops. The counts of all of these species did not differ between conventional and GMHT treatments in any of three crops studied (beet, maize and spring oilseed rape; Haughton *et al.*, 2003). Dominance in spring oilseed rape was the only significant treatment effect observed (i.e. the abundance of carabids was greater

under GMHT cropping). However, counts of Carabids that feed on weed seeds were reduced in GMHT crops (Brooks *et al.*, 2003), indicating a possible indirect effect due to the decrease of weed populations.

True bugs (Heteroptera)

Predatory heteropterans are common and important members of the natural enemy complex of a variety of row crops (Schaefer and Panizzi, 2000). Moreover in a field study performed in Switzerland this group of insects showed the highest correlation with the estimate of overall biodiversity in an agricultural landscape (Duelli and Obrist, 1998). The Cry3Bb1 protein at a concentration ten times higher than expected in Bt maize had no adverse effect on the survival and development of Orius insidiosus Say nymphs in experiments performed by Duan et al. (2008). Rauschen et al. (2009) examined the impact of Bt maize on the mirid Trigonotylus caelestialium (Kirkaldy) in field experiments and concluded that there was no evidence for a negative impact of MON88017 maize on the abundance and body weight of this species, despite its considerable exposure to Cry3Bb1. Torres and Ruberson (2008) assessed the effects of Crv1Ac-expressing cotton on several predatory heteropterans in caged plants experiments. They concluded that the toxin was conveyed to predators via their prey (different species were used), and that there were no effects on the life history traits of the predators (i.e. pre-imaginal development, body weight, reproduction and survivorship). For instance, Podisus maculiventris Say exhibited similar life history characteristics (developmental time, survival, longevity, and fecundity) when preying upon Bt or non-Bt cotton fed Spodoptera exigua (Hubner) larvae (Torres and Ruberson, 2008). Arpaia et al. (2007) reported the results of a three-year study of several predatory guilds in experimental plots of Cry3Bb-expressing aubergine and their near isogenic lines. Mirids were abundant in these experimental fields and no differences between treatments were detectable. Some species (e.g. Macrolophus caliginosus Wagner, Dyciphus errans – Wolff–) were significantly associated with GM aubergine areas on a few of the sampling dates.

3.1.2. Parasitoids

Parasitoids can also be exposed to metabolites expressed in GMPs via one or more trophic levels (e.g. direct feeding on GM plant material, mainly nectar or exudates, or by their host organisms feeding on GMP tissues). When reviewing the available body of data, invertebrate parasitoids appeared to be more sensitive than predators to diets containing Cry toxins or proteinase inhibitors under laboratory conditions (Lövei *et al.*, 2009); in fact 25 out of 31 response classes evaluated in this analysis were significantly non-random. The meta-analysis conducted by Naranjo

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(2009) confirmed the higher sensitivity of parasitoids to Cry toxins. In addition, by evaluating tri-trophic and bitrophic experiments separately, this author was able to emphasise that host quality is highly relevant in determining such results. In fact, mostly negative effects occurred when parasitoids were feeding on hosts sensitive to the toxins expressed in GMPs. The intimate relationships of these natural enemies with their hosts renders them sensitive to any change in host guality (Godfray, 1994). However, indirect effects do not fully explain the published results, and other possible mechanisms need to be considered. Ramirez-Romero et al. (2007) observed that exposure to Cry1Ab toxin via hosts fed Bt maize tissue sub-lethally affected the parasitoid wasp Cotesia marginiventris (Cresson). In experiments where the performance of this parasitoid developing on aphids fed MON810 maize was compared with those on aphids fed control maize (which were similar in size), negative effects were detected with respect to the wasp developmental times, adult size, and fecundity. Conversely, these negative effects were not observed when toxin-containing artificial diet was used in tritrophic experiments; the authors were thus able to demonstrate the importance of the plant as a medium in causing negative effects at the third trophic level.

In some cases, an enhanced performance of parasitoids was observed when their hosts were obtained from Cry-expressing plants. For instance, Schuler et al. (2003) showed that the number of emerging Cotesia plutellae (Kurdjumov) adults was higher on Cry1Ac-expressing oilseed rape compared to control plants. Faria et al. (2007) detected a positive effect of six varieties of Bt maize (including events Bt11, MON810 and Bt176) on the performance of the maize leaf aphid Rhopalosiphum maidis (Fitch) in growth chamber experiments, which in turn enhanced the performance of the parasitic wasp C. marginiventris that also utilised the aphid honeydew. In these experiments, GM maize lines were generally significantly more susceptible to aphids than their near-isogenic controls. The analysis of plant sap in selected maize lines revealed significantly higher amino acid levels in Bt maize, which might partially explain the observed increased aphid performance. Larger colony densities of aphids on Bt plants resulted in an increased production of honeydew that is normally used as food by beneficial insects. In fact, C. marginiventris females lived longer and parasitised more pest caterpillars in the presence of aphid-infested Bt maize than in the presence of aphid-infested near isogenic maize.

Reports from field studies have highlighted a diminished number of parasitoids (reviewed in Wolfenbarger *et al.*, 2008) in areas with *Bt*-expressing maize compared to respective untreated controls. The author of this review showed that

this pattern was entirely explained by the reduced abundance of *Macrocentris* grandii Goidanich, an exotic specialist parasitoid of the European corn borer, and therefore the absence of the natural host was thought to be the cause of the differences found in maize fields. However, these specific indirect effects need to be carefully considered in an ecological context, e.g. abundance of alternative hosts for the parasitoids in the field and adjacent areas, abundance and diversity of parasitoids for the pest and non-pest species linked to the crop etc. (Arpaia *et al.*, 2006a), in order to evaluate the possible repercussions on this functional guild.

3.2. Pollination

Many crop plants, including crop species that have been genetically modified to date, are dependent upon pollinators for optimal reproduction. In some cases, crop yield is strictly dependent upon pollinators activity in the field (e.g. watermelon, squash, kiwifruit) or in the greenhouse (e.g. tomatoes, strawberries), whilst in other cases, the effective presence of pollinators allows for greater and more stable yields (e.g. oilseed rape, soya bean, cotton; Klein et al., 2007). Some of these crops are also important for honey production in many countries (Crane and Walker, 1986; Free, 1993). The successful establishment of mutualistic interactions between plants and pollinators is dependent upon a series of successive steps that require a finely tuned combination between plant attractants and insect senses. Flowers announce their rewards (most commonly pollen and nectar) to pollinators using colour, scent, size, or shape, making it easier for visitors to recognise them. From an insect perspective, smell and vision are therefore essential for efficiently finding food sources. Pollination ecology plays a fundamental role in protecting biodiversity, and natural selection has resulted in efficient pollinating species to out-compete other species for the same food source.

A recent review on the possible effects of GMPs on pollinators (Malone and Burgess, 2009) concluded that none of the commercially available GM crops expressing herbicide tolerance or insect resistance traits have deleterious impacts on pollinators. The only *Bt* toxin known to be specifically active against Hymenoptera (the order which includes most of the important pollinating insects) is Cry5, which functions with a similar mechanism to Cry1 toxins (Garcia-Robles *et al.*, 2001). No Cry5-expressing GMPs are currently commercially available. Feeding studies performed in controlled conditions with honeybees fed either with *Bt* pollen or mixtures of honey or sugar syrup containing purified Cry1Ab toxin have indicated no direct adverse effects on larvae and adult survival (Ramirez-Romero *et al.*, 2005, 2008; Rose *et al.*, 2007). These findings concur with earlier studies conducted with Cry3B under semi-field conditions (Arpaia, 1996). Based on a meta-analysis of 25

independent laboratory studies assessing direct effects on honeybee survival of Cry toxins from currently commercialised *Bt* crops, Duan *et al.* (2008) concluded that Cry toxins do not negatively affect the survival of either honeybee larvae or adults in laboratory settings. Nevertheless, the same authors considered that in field settings, honeybees might face additional stresses, which could theoretically affect their susceptibility to Cry toxins and generate indirect effects.

Some studies focused on the development of the hypopharyngeal gland in honeybees. Hypopharyngeal glands are considered an important indicator of bee life history, as they are used in nurse bees to prepare brood food (jelly) for the larvae. In this respect, Babendreier et al. (2005) fed young adult bees for 10 days with Bt maize pollen expressing Crv1Ab toxin (event MON810) or with purified Cry1Ab toxin in sugar solutions. No significant differences in diameter, weight and development of hypopharyngeal glands were found in bees fed either Bt pollen or *Bt*-containing sugar solutions, compared to their respective controls. In a field study where colonies foraged on Cry1Ab-expressing maize (event Bt11) and were fed Bt pollen cakes for 28 days, Rose et al. (2007) did not observe adverse effects on bee weight, foraging activity, and colony performance. Similarly, in a flight cage study in controlled conditions, no significant differences were reported in honeybee mortality, syrup consumption and olfactory learning performance when honeybee colonies were exposed to different syrups containing Crv1Ab protoxin (Ramirez-Romero et al., 2005). In a later paper, Ramirez-Romero et al. (2008) state that the toxin did not cause lethal effects on honey bees; however, feeding behaviour was affected when exposed to very high concentrations of Cry1Ab (5000 ppb), with honey bees taking longer to imbibe the contaminated syrup. The authors state though, that these negative effects are unlikely in natural conditions, since exposure to such large amounts of toxin is not foreseeable.

Other insect resistant traits are possibly more hazardous to bees. Serine proteinase inhibitors may affect honeybees and bumblebees at high concentrations while cisteine-based proteinase inhibitors do not seem to have similar effect (Malone and Burgess, 2009). The exception is aprotinin, a serine proteinase inhibitor which affects honeybees but not bumblebees (Burgess *et al.*, 1996; Malone *et al.*, 2000). In the case of herbicide tolerant GM plants, possible negative effects may arise due to changes in cultivation practices which lead to altered weed composition. Reduced flowering in the agro-ecosystem may indirectly lead to a lower local abundance of pollinators (e.g. Haughton *et al.*, 2003). An analysis of the environmental impact of GM plants, if based only on acute toxic response, might prove insufficient to draw general conclusions about their safety for pollinators. In analysing the case study

of *Bt* cotton in Brazil, Arpaia *et al.* (2006b) suggested that other possible adverseeffect scenarios should be duly considered at both the organism and colony level. Amongst these, any possible impaired colony development via the lowering of queen fecundity or any modification of foraging behaviour due to an impaired ability to find food may have important consequences on overall pollination activity in the agro-ecosystem.

The exposure of pollinator hymenoptera to toxins expressed in GM plants is a direct consequence of their feeding habits, since these organisms occupy the second trophic level in food chains. Constitutive promoters in transgene constructs generally lead to limited protein expression in pollen (Potenza et al., 2004). In some events though, significant amounts of Cry toxin is expressed in pollen (e.g. Koziel et al., 1993; Mattila et al., 2005; Yao et al., 2006). However, the presence of newly-expressed proteins in GM plant nectar is considered negligible since it has no cellular content (Malone and Burgess, 2009). The GMP species will also modulate the possible exposure of bees to toxins; for instance, maize does not represent a major pollen source for bees. Babendreier et al. (2004) reported that fully grown worker bee larvae contain between 1720 and 2310 maize pollen grains in their gut before defecation, corresponding to 1.52-2.04 mg of pollen consumed per larva. Adult honeybees consume significant quantities of pollen while pollen consumption by larvae is minimal, especially during early growth stages, and therefore larvae are less exposed to GM plant-produced toxins. By directly feeding larvae with pollen, Babendreier et al. (2004) calculated the contribution of Cry protein to be less than 5 % in relation to the total amount of protein necessary for complete larval development. Moreover, due to the low concentration of Cry1Ab in pollen, honeybees will only be exposed to low concentrations of proteins. The presence of pollen in honey reserves in the hives can prolong the exposure of the colony to toxins that are expressed in pollen grains and therefore the time of flowering does not fully represent the extent of exposure these social bees may be faced to. Pierre et al. (2003) observed the diversity and density of the foraging insect population on GM HT OILSEED RAPE genotypes to be similar to that on the near isogenic controls. Moreover, the foraging behaviour strategy was similar between treatments.

3.3. Biological Components of Soil Fertility

Soil fertility can be defined as the characteristics (biological, physical, chemical, etc.) enabling a soil to provide nutritive elements that support plant growth, and its production in agricultural fields. While it is known that physical, chemical and biological components of soil fertility strongly interact, in this context mainly

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the latter will be considered. The availability of nutrients is strongly dependent on effective microbial functioning in the soil. Communities of soil-dwelling invertebrates are also involved in nutrient cycling and decomposition of organic matter that have both an ecological and practical significance for agriculture (Moore *et al.*, 1988). Species diversity in soils is extensive (e.g. Curtis *et al.*, 2002), as soil-dwelling organisms can account for most of the whole agro-ecosystem diversity. Plants themselves, through the release of the exudates in the soil, exert a major influence in selecting communities of micro-, meso- and macroorganisms in the rhizosphere. Brussaard *et al.* (2007) suggest that soil biodiversity can be assessed, managed and conserved, showing examples of successful and unsuccessful practices which have been used in various regions of the world to manage soil biodiversity.

In the context of this review, it is recognised that soil fertility is another important ecosystem service guaranteed to farmers through the establishment of intricate food webs (both grazing and detritus food webs are guite active in agricultural soils). Potential adverse environmental impacts due to the cultivation of GMPs may occur should soil organisms be negatively impacted and/or changes in microbe-mediated functions in the soil occur (Icoz and Stotzky, 2008). GMPs can expose non-target soil-dwelling organisms to proteins via root exudation and cultivation residues. The persistence of toxins in the soil can be guite variable, and is particularly dependent upon soil type; for instance, Cry toxins are known to bind to clay, suggesting that there is potential for their long term persistence and consequently long exposure to non-target organisms (Icoz and Stotzky, 2008). Commonly, experiments have shown GMPs to exert changes in the local populations of soil organisms. Generally speaking though, these differences have been considered to be in line with those effects induced by changing crop species or even varieties within a single species (EFSA, 2009). It is therefore important that both statistical significance and biological relevance are considered when assessing possible specific environmental impacts.

This section will mainly consider the available information relating to GMPs and soil arthropods, and then will briefly summarise the state of the art and the open issues related to the possible effects on soil microorganisms; while for more detailed information on effects on microorganisms, interested readers are recommended to consult more specific literature.

3.3.1. Earthworms

Earthworms play an important role in decomposing plant litter, and are responsible

for numerous physical changes that affect the biological properties and processes in soil (e.g., through the maintenance of soil structure) and are considered important organisms in the regulation of nutrient cycling processes (Icoz and Stotzky, 2008). Laboratory studies performed on some earthworm species, such as *Aporrectodea caliginosa* Savigny (Vercesi *et al.*, 2006; Schrader *et al.*, 2008), *Eisenia foetida* Savigny (Clark and Coats, 2006) and *Lumbricus terrestris* L. (Saxena and Stotzky, 2001a; Zwahlen *et al.*, 2003; Lang *et al.*, 2006; Schrader *et al.*, 2008) did not reveal significant adverse effects on earthworm survival, growth and reproduction upon ingestion of Cry1Ab toxin.

No adverse effects on mortality or weight were observed on *L. terrestris* exposed to soil incorporating plant material from *Bt*-expressing maize after 40 or 45 days, respectively, compared to non-Bt maize (Saxena and Stoztky, 2001a). Zwahlen et al. (2003) investigated L. terrestris mortality and growth in laboratory and field experiments by exposing juveniles and adults to Bt11 maize (expressing a Cry1Ab toxin) for up to 200 days. Even though earthworms were not lethally affected by the exposure to Bt maize, sublethal long-term effects were observed in the laboratory study: the growth of adults, expressed as mean fresh weight, was similar for 160 days, but significantly declined thereafter in Bt-exposed earthworms. However, in the field no adverse effects of Bt maize were found (Zwahlen et al., 2003). Laboratory studies by Clark and Coats (2006), in which E. foetida was fed leaf material from either Bt maize (events Bt11 and MON810) or the near isogenic counterpart in a soil system did not reveal adverse effects on survival or reproduction after 4 weeks. Vercesi et al. (2006) studied the effects of MON810 maize on important life-history traits (survival, reproduction and growth) of A. caliginosa under various experimental conditions. The authors measured the growth of juveniles until maturity, as well as cocoon production and hatchability. Finely-ground leaves of MON810 maize added to soil had no adverse effects on these life-history traits in A. caliginosa, even when exposed to very high concentrations. In addition, the growth of juveniles was unaffected when they were kept in pots with *Bt*-expressing maize plants for 4 weeks. A statistically significant negative effect of high concentration of Bt maize residues was only observed on cocoon hatchability. However, since earthworms were exposed to very high concentration of fresh Btexpressing plant material in the experiment, the authors questioned whether the negative effect would have any ecological significance under field conditions. In recent experiments performed by Schrader et al. (2008), A. caliginosa and L. terrestris both survived incubation in microcosms for 5

weeks, irrespective of whether they grew in the presence of leaves and roots of MON810 or control maize. The content of Cry1Ab toxin detected in the plant residues strongly declined over time.

Saxena and Stotzky (2001a) demonstrated the uptake of Cry1Ab toxin by earthworms via detection in casts and guts. The ingestion of Cry1Ab toxin by earthworms was later confirmed through the detection of the protein in their gut and faeces (Zwahlen *et al.*, 2003). Field surveys indicated no adverse effects on earthworms during the cultivation of *Bt* maize expressing the Cry1Ab toxin (e.g., Zwahlen *et al.*, 2003; Krogh *et al.*, 2007). No significant differences were reported in the population density or total biomass of *Lumbricidae* between soils with *Bt* (events MON810 and Bt176) and non-*Bt* maize at 5 sites during 4 years of maize field cultivation, though the site and the year had a significant influence on both measurement endpoints (Lang *et al.*, 2006).

3.3.2. Nematodes

The composition of nematode communities is considered an useful bioindicator of soil health because it correlates well with nitrogen cycling and decomposition, two critical ecological processes in soil (Neher, 2001). Nematode abundance and diversity has been investigated in several experiments with GMPs; in experimental conditions, these values varied substantially when GMPs were compared to near isogenic controls. The extent and the direction of this variability however, is not unequivocal.

Cry toxins have been proposed to be hazardous to the nematode species *Caenorhabditis elegans* (Maupas) following laboratory studies by Höss et *al.* (2008). The authors studied the potential toxic effects of Cry1Ab toxin on *C. elegans*, either by exposing the species to rhizosphere and bulk soil from experimental fields cultivated with *Bt* maize (event MON810) or to different solutions of Cry1Ab toxin expressed in *Escherichia coli*. Nematode reproduction and growth were significantly reduced in rhizosphere and bulk soil of *Bt* maize as compared with soil from isogenic maize, and were significantly correlated with the Cry1Ab toxin concentrations in the soil samples. However, according to toxicity tests with the pure Cry1Ab protein, it was determined that the concentrations of Cry1Ab toxin measured in the *Bt* maize soil samples were not sufficiently high to produce direct toxic effects on *C. elegans*. Therefore the authors suggest that the adverse effects. In addition, the possibility of interactions of the newly-expressed toxin with other compounds produced by the GM plant should also be considered

as a possible mechanisms to explain these results. Saxena and Stotzky (2001a) found no significant differences in the number of nematodes in the rhizosphere of Bt and non-Bt maize (event NK4640Bt) in growth chamber studies. In a glasshouse study, Griffiths et al. (2006) reported significantly higher nematode populations of Acrobeloides spp. and Pratylenchus spp. when grown in the presence of Bt maize (event MON810) than with non-Bt maize. Further, in a later glasshouse study involving 8 different varieties of maize Bt, Griffiths et al. (2007b) went on to report that nematode abundance varied mainly between maize varieties, rather than between Bt and non-Bt maize, and commented that differences in previously published studies on soil nematodes in Bt maize were smaller than varietal effects. Field experiments confirmed that changes to nematode communities due to Bt maize (event MON810) were small and transient, and smaller than those induced by seasonal, soil type, tillage, crop type or varietal effects (Griffiths et al., 2007a). Effects of Bt maize (events MON810 and Bt176) on the plant pest Pratylenchus spp. and the bacteriovorius C. elegans, have also been studied in field trials in Germany (Lang et al., 2006). No adverse effects of GM maize were observed with respect to Pratylenchus spp. population density, whilst conversely C. elegans growth, number of eggs and reproduction rate were negatively affected. Further studies by Höss et al. (2008) confirmed those findings.

Maize does not represent the only GM crop studied for possible effects on nematodes. Manachini *et al.* (2004) studied nematode species assemblage in experimental fields with oilseed rape expressing Cry1Ac toxin. The authors found a statistically significant shift in community structure, and a direct correlation between GM oilseed rape and the abundance of fungal feeding nematodes, while phytophagous nematodes were significantly associated with near isogenic areas. These results were in agreement with similar surveys conducted in Northern Italy (Manachini and Lozzia, 2002) where nematodes feeding on fungi were more abundant in *Bt* maize fields. A possible explanation for the presence of differences in plant composition that are known to occur in some GMPs (e.g. lignin in maize, Saxena & Szotzky 2001b) which could in turn have effects on the detritus food webs in the soil.

3.3.3. Isopods

The woodlouse (*Porcellio scaber* Latreille) is considered a model decomposer organism, and has been used in laboratory feeding studies for detecting potential adverse impacts of Cry1Ab toxin and Cry1Ab-expressing maize. Assimilation of

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the Cry1Ab toxin by P. scaber following intentional exposure was demonstrated by the detection of the toxin in the faeces after consuming *Bt*-expressing plant material (Wandeler et al., 2002; Pont and Nentwig, 2005). The latter authors observed that part of the Bt toxin taken up by primary decomposers is not digested and therefore is released in its active form into the soils. Under field conditions in autumn and winter, the toxin remains active and available to soil organisms until the next field season. No adverse effects on P. scaber consumption, survival and growth were observed when the species was fed plant material from maize expressing Cry1Ab toxin (Escher et al., 2000). The survival and growth of two other common isopod species, Trachelipus rathkii (Brandt) and Armadillidium nasatum Budde-Lund, were not adversely affected after exposure to the purified Cry1Ab toxin or leaves of different Bt maize events under laboratory conditions for 8 weeks (Clark et al., 2006). In other experiments however, some differences in mortality, weight gain and consumption by isopods and in the digestibility of plant material were detected. In these studies (Wandeler et al., 2002; Clark et al., 2006), the differences in the composition and nutritional guality of maize varieties used possibly contributed to determine the differences between treatments, and it is therefore difficult to draw conclusions about the effective consequences from the transgene into the test system.

3.3.4. Collembola

Collembola are an important group of organisms involved in the breakdown and recycling of crop residues, and they are normally abundant in agricultural fields (Hopkins, 2006). They often live in the root zone of plants and can therefore be exposed to root exudates and residues (Icoz and Stotzky, 2008). As such, Collembola are considered good indicator species of soil fertility and health, and have been used to detect the potential impact of Cry toxins. A study adding 4 purified Bt insecticidal proteins (Cry1Ab, Cry1Ac, Cry2A, and Cry3A) at concentrations of 200 mg/g to the diet of Folsomia candida Willem and Xenylla grisea Axelson for 3 weeks did not detect any impacts on the survival or reproduction of the study species when compared with a control diet (Sims and Martin, 1997). No deleterious effects on F. candida survival and reproduction were observed when fed leaves of Bt maize expressing the Cry1Ab toxin compared with those fed leaves of the isolines (Clark and Coats, 2006). Bakonyi et al. (2006) showed that Bt maize (event MON810) was less preferred as a food by F. candida than was near-isogenic control maize, while this preference was not observed for Heteromurus nitidus (Templeton) and Sinella coeca (Schott). During these experiments, F. candida defecated 30 % less around Bt maize in choice tests, but did not show a preference to remain on either of the two plant material types offered (*Bt* and non-*Bt* maize). When starved individuals were tested in the same choice experiments, they equally consumed both diets. In addition to the presence of the Cry1Ab toxin, available diets also differed in C/N ratios in respective plant material. Heckmann *et al.* (2006) reported that the growth and reproduction of *Protaphorura armata* (Tullberg) reared on roots of *Bt* maize expressing Cry1Ab toxin were not significantly different from those reared on roots of non-*Bt* maize for 4 weeks. *P. armata* performed significantly better on a diet of yeast amended with purified Cry1Ab toxin than on root tissue of *Bt*- and non-*Bt*-expressing maize. In field studies, no significant differences in the population density of collembolans were found in soils cultivated with *Bt*- and non-*Bt*-expressing maize (Lang *et al.*, 2006). Concentrations of Cry toxins in plant material in soils in the field are usually low and estimated to be less than 30 µg/g of fresh weight, suggesting that these concentrations should not pose a relevant threat to Collembola (Sims and Martin, 1997).

3.3.5. Other taxonomic groups

Even though diplopods are not the most important group of decomposers of plant litter in soil, they are widely spread in the agricultural landscape and regularly occur in maize fields (EFSA, 2009). Laboratory studies have been performed on Allajulus latestriatus (Curtis) to analyse the effects of Bt maize (event Bt11) on mortality, food consumption, weight gain, and faeces production. No significant differences were found when test organisms where fed Bt maize compared to the near isogenic control and two commercial varieties used as references. Exposure to very high Cry1Ab concentrations (more than 100 times higher than those detectable in leaves) did not result in a significantly higher mortality (Weber and Nentwig, 2006), Faeces production was significantly increased when animals were maintained on Bt maize. Diplopods prefer feeding on partly degraded plant material which usually contains lower Cry1Ab concentrations (Weber and Nentwig, 2006), therefore their exposure in field conditions is estimated to be rather low. A recent paper (Hönemann and Nentwig, 2009) described laboratory feeding studies using GMPs and the enchytraeid Enchytraeus albidus Henle. Through their feeding activities, enchytraeid worms support mineralisation processes and improve the structure of soil. As for other soil-dwelling arthropods, enchytraeids can be exposed to the toxins released over time in the soils by GMP exudates or residues. Enchytraeids were observed to feed on diets that contained leaf material of Bt maize, though no Cry1Ab toxin was detected in adults after 3 weeks of feeding (Hönemann and Nentwig, 2009). In addition, E. albidus survival and reproduction showed no significant differences between the Cry3Bb1-expressing treatment and the untransformed counterpart. Different results were obtained

when Cry1Ab toxin was part of the diets of the enchytraeid species. Significantly more individuals were reported to survive in the treatment with Bt11 maize than with the corresponding near-isogenic control. In contrast, a significantly higher number of offspring were generated in the control treatment, as compared with the Bt11-containing diet. Due to differences in plant composition between the maize varieties, the authors could not tease out how much the presence of *Bt* maize leaf material in their diet contributed to such differences. Generally, enchytraeid worms do not feed on a single food source, but take up all degradable organic matter of adequate size in the field and therefore their exposure to toxins is likely to be reduced in normal field conditions (EFSA, 2009).

3.3.6. Effects on soil microorganisms

Microorganisms are the most abundant organisms in soils and are involved in many fundamental processes (e.g. decomposition of organic matter, mineralisation, decomposition of chemicals, improvement of soil structure, etc.; Gupta and Yeates, 1997). Root exudates released by plants selectively regulate which organisms reside in the respective rhizosphere (e.g. Lynch, 1994). Therefore, qualitative and quantitative changes in their emission could influence the diversity and activity of soil microbiota (Icoz and Stotzky, 2008). Populations of soil microorganisms are known to be affected by many factors in field conditions, therefore isolating the possible effects of a new stressor requires accurate experimental plans, moreover not all soil microorganisms can be easily grown in the laboratory and therefore experiments in controlled conditions can not always be easily performed.

While the majority of the studies summarised by Icoz and Stoztky (2008) indicated that *Bt*-expressing plants cause no or minor changes in microbioal communities, in some studies relevant differences in the presence of microorganisms between soils cultivated with *Bt* and non-*Bt* maize were demonstrated. Xue *et al.* (2005) found a lower ratio between gram-positive and gram-negative bacteria in soil with *Bt* maize compared to their controls, while the effect was reversed in *Bt*-expressing potato. Root exudates of *Bt* maize (event Bt176) were shown to reduce pre-symbiotic hyphal growth of the arbuscular mycorrhizal fungus *Glomus mosseae*, as compared with those of another *Bt* maize (event Bt11) and control maize (Turrini *et al.*, 2004). Castaldini *et al.* (2005) also reported consistent differences in rhizosphere heterotrophic bacteria and mycorrhizal colonization (including *G. mosseae*) between *Bt* maize (event Bt176) and its conventional counterpart. According to the authors, the genetic modification in Bt176 maize might have led to changes in plant physiology and composition of root exudates, which in turn may have affected symbiotic and rhizosphere microorganisms. For

instance, microbial activity could have been affected by soluble sugar content (Burns and Dick, 2002); percentage differences in sugar content in the plants used in the study were relatively high and this could have also contributed to the observed changes. From an ecological perspective, Widmer (2007) suggests that effects observed on symbiotic microorganisms will only be disadvantageous for the crop itself, without representing a concern for the ecosystem. A more recent field study (Knox *et al.*, 2008) reported very different results when the production of arburscural mycorrhizae was studied in GM cotton plants. The experiments included commercial cultivars of cotton expressing genes for insect resistance (Cry1Ac and Cry2Ab), glyphosate tolerance (5-enolpyruvylshikimate-3-phosphate synthase gene), or both, and their conventional parent lines. The development of mycorrhizae in cotton roots increased rapidly in the first three weeks after sowing and the pattern of colonisation was virtually identical among both conventional and GM cultivars of cotton at each assessment.

Reported effects on microbial communities were in general considered spatially and temporally limited, and small compared with those induced by differences in geographic location, temperature, seasonality, plant variety and soil type (Fang *et al.*, 2005, 2007; Griffiths *et al.*, 2005, 2006; Lilley *et al.*, 2006; Filion, 2008; Icoz and Stotzky, 2008). Factors such as plant growth stage and field heterogeneity produced larger effects on soil microbial community structure than MON810 maize (Baumgarte and Tebbe, 2005; Griffiths *et al.*, 2007b). However, some issues in this area are still to be resolved. For instance, in some cases it is not clear if the methodology used for analysing soil samples has an influence in the appearance of diverging results obtained for similar groups of microorganisms in the literature. Moreover, the relationship between the high biodegradability and adsorption of Cry toxins in the soil and their biological activity should be further clarified. However, a more detailed discussion in this area goes beyond the goal of the present review.

3.4. Measuring the Overall Functionality

It should be apparent to the reader by now that selecting the appropriate assessment endpoint(s) is very important in analysing any non-target effects of GM crops. In line with the suggested "functional approach", the choice of one or a few species amongst each functional group is a sensible way of studying possible environmental impacts. However, in some cases, the preliminary selection of indicator species for the risk assessment of GMPs may be difficult to the extent that it might not even be considered as the most appropriate approach. In that case, an alternative or complementary strategy could be the estimate of the functionality of a given ecosystem service.

In a field study on the arthropod fauna in Cry1Ac-expressing cotton, Naranjo (2005a) demonstrated that the abundance of some predator species was significantly diminished in plots with GMPs compared to those with near-isogenic untransformed cotton plants. However, in a parallel study conducted in the same cotton fields, the author found that the overall predation rate on the pink bollworm Pectinophora gossypiella (Saunders), one of the target species of Cry1Ac cotton, was similar between control and GMP plots (Naranjo, 2005b). Therefore, the detected changes in the guilds of predators did not impair the ecosystem service of natural predation on this pest. In this case, the consideration of only a given predator-prey dynamics might have resulted in the wrong conclusions. In two field studies conducted with GM potato and aubergine (and their respective untransformed controls), Arpaia et al. (2009) observed that the predation rates on L. decemlineata Say egg masses due to a guild of predatory species were similar in GM and control plots. In the two different experiments, L. decemlineata was the potential target for GM aubergine expressing the Cry3Bb toxin, while it was a non-target species for GM potato which were resistant to Lepidoptera by expressing a Cry1Ab toxin. The fact that the eggs were preyed might have limited the predator exposure to the Cry toxins in all cases, since the eggs deposited on leaves are not likely to absorb the toxin from plants. These results enabled the authors to conclude that the ecological function of natural predation on L. decemlineata eggs in GM plots was not impaired.

An indirect measure of pollination activity (e.g. by comparatively measuring fruit set between pollinated GM and control crops) might furnish a general indication of the foraging activity of pollinators in this type of experiment. Studies in which the decomposition of *Bt* maize was compared with that of non-*Bt* isogenic lines mostly showed that Cry1Ab-expressing maize did not affect decomposition rates or the mass of carbon remaining over time (e.g. Cortet *et al.*, 2006; Tarkalson *et al.*, 2008). Likewise, litter-bag experiments with *Bt* maize (event Bt11) reported by Zwahlen *et al.* (2007) did not reveal major changes in the decomposition rate of *Bt*-maize residues. Van Toan *et al.* (2008), discussing a possible risk assessment strategy for *Bt* cotton in Viet Nam, state that the complexity of soil ecosystems render studies based on species lists impractical and unreliable. The authors suggest that ecosystem processes such as biomass decomposition, cellulose and lignin breakdown, phosphorous and nutrient uptake should constitute the focus of the biosafety studies.

These examples show that on a case-by-case manner, different assessment endpoints can be effectively chosen in line with the proposed functional approach and therefore risk assessors should evaluate this possibility considering the combination of plant, newly expressed trait(s), and the receiving environment. An additional criterion, while not strictly based on ecological characteristics, that may help to assess an environmental impact on NTOs is the consideration of anthropocentric values. This criterion will possibly lead to the consideration of other NTOs that could not be selected among the above-mentioned functional groups. Anthropocentric values are related to human goals and require the consideration of organisms characterised as secondary pest species, rare or endangered species, species that generate income, and species of social or cultural value (Birch et *al.*, 2004).

A very well known case of risk assessment of GMPs for a non-target species concerns the Monarch butterfly D. plexippus L., a species which can not be ascribed to any functional group of NTOs directly involved in ecosystem services. This lepidopteran species though has an important charismatic value, especially in the USA, due to its fascinating habits, especially its periodical massive migrations across the continent. The report of the hazard presented to D. plexippus larvae from the ingestion of Cry1Ab-expressing maize pollen in laboratory experiments (Losey et al., 1999) triggered extensive laboratory and field studies to determine whether Monarch butterfly populations would be at risk under realistic exposure conditions from *Bt*-expressing maize pollen in the USA. Monarch larvae do not directly feed on maize leaves, therefore pollen deposition on their host plants, the milkweed Asclepias syriaca L., is the most realistic exposure route for the larvae to Cry toxins expressed in maize. Field studies indicated that the overall proportion of Monarch butterfly populations exposed to toxic levels of *Bt* pollen was small due to the limited spatial distribution of pollen (Pleasants et al., 2001) and the limited temporal overlap between larval development and pollen shed (Oberhauser et al., 2001). Based on the USA data, a risk assessment model estimated that 50 % of the breeding population of the Monarch butterfly was potentially exposed to Cry1Ab-expressing pollen in the USA corn belt (Sears et al., 2001), but that only an additional 0.6-2.5 % mortality would be generated due directly to the cultivation of Bt-expressing maize (Dively et al., 2004). Anderson et al. (2004) and Prasifka et al. (2007) both reported a reduction in feeding and weight gain of the *D. plexippus* larvae under laboratory conditions due to food induced behavioural changes, but how these results would correlate with feeding habits in the field was not elucidated. Under field conditions early instars, the most susceptible to the Cry1Ab toxin, are less exposed to Bt pollen drift, as they mainly feed on the upper third of milkweed plants where the lowest

densities of anthers occur (Pleasants *et al.*, 2001; Anderson *et al.*, 2004). In addition, larvae can move to the underside of leaves where they would avoid any contact with anthers (Pleasants *et al.*, 2001; Jesse and Obrycki, 2003).

Maize plants expressing lepidopteran-specific Cry toxins were shown to be hazardous to a range of other Lepidopteran species, some of which are species of conservation concern (e.g. Felke et al., 2002; Lang and Voitech, 2006). These species could also be exposed to potentially toxic pollen deposited on their host plant in and around maize fields. The possible effects on non-target Lepidoptera has been invoked as a justification for several safeguard clauses that were risen by several European nations (e.g. Austria, France, Greece, Germany, Hungary, Luxembourg) who asked for the ban of MON810 maize cultivation in their territories. The European Food Safety Authority GMO Panel (EFSA, 2009) conducted an exposure assessment based on a simulation model to help quantify the risk assessment. Exposure was modelled for 3 combinations of lepidopteran species and their host plants, all of which occur widely throughout the European Union. These were: the butterflies Inachis io L. and Vanessa atalanta L. both feeding on the host plant Urtica dioica L.; and the pest species Plutella xylostella L. (diamondback moth) and its host plant species in the Brassicaceae family. The analysis was based on an 11-parameter deterministic mathematical model, of which 7 parameters were specific to particular geographic regions and 4 parameters were more generic to the particular species/host plant combination. For the majority of areas where U. dioica is known to occur, for both the butterfly species considered, the best estimate for mortality was less than one individual in every 1800, whilst sub-lethality was estimated at less than one individual in every 550. When the diamondback moth was considered, for the majority of areas the best estimate for mortality was less than one individual in every 300, whilst sub-lethality was less than one individual in every 100 (EFSA, 2009). A further elaboration of this mathematical model (Perry et al., 2010), which includes more experimental values for two of the estimated parameters and accounts for the possibility of aggregated pollen dispersion, suggests that the probability of sublethal effects might be four times less than previously estimated. In order to obtain these estimates, extrapolations were made about the toxicity of MON810 maize pollen in relation to other Bt maize events with different toxin expression in pollen. It has become clear that a case-by-case evaluation will be necessary. This will have to consider the biology of NTOs in relation to the specificity of the receiving environment, and also the specific GMP and its molecular characteristics (including toxin expression in different plant parts).

3.5. What Else is Non-Target?

Agro-ecosystems and adjacent habitats are the first environments to be exposed to GMPs and their products, and therefore they are correctly considered in risk assessments. Nevertheless, the spatial-temporal dispersion of GM products can influence other trophic chains that in specific cases might become relevant. For example, pollen dispersal might bring other herbivore species in contact to newly-expressed toxins at distances from crop fields; pollen and plant residues may enter water bodies surrounding cultivated fields and therefore enlarge the possible exposure to expressed toxins. Trophic chains do not stop with predatory arthropods, such as spiders, but commonly involve other taxa, for instance, birds. Finally it must be considered that in some conditions non-arthropod herbivores (e.g. snails, rodents, wildlife) may commonly feed on cultivated plants.

The water flea Daphnia magna Straus is generally considered a surrogate species for assessing the effects of pollutants on aquatic organisms, and as such, the species is commonly included in eco-toxicological tests performed by the applicant when submitting dossiers for the commercialisation of GM crops or products. It has been reported that the species did not suffer acute toxic effects in laboratory studies with Cry1Ab toxin (Mendelson et al., 2003). However, in contradiction, a laboratory experiment performed by Bøhn et al. (2008) revealed that D. magna fed with a suspension of MON810 maize flour had a higher mortality and lower proportion of females reaching sexual maturity. Since maize flour is not a common natural diet of flea beetles, and that D. magna fed non-Bt maize also experienced delays in development, the experiment could not rule out the possibility that these results might have been caused by nutritional deficiencies related to the maize-based diet. Rosi-Marshall et al. (2007) reported that by-products of Cry1Ab-expressing maize entered headwater streams. In laboratory experiments, the authors found reduced growth and increased mortality in aquatic Trichoptera; nevertheless, no effects were found during samplings in natural conditions by Chambers et al. (2007). The study by Rosi-Marshal et al. (2007) has attracted a lot of controversy especially with respect to its experimental design, but it did highlight a potential hazard of high doses of Cry1Ab toxin for Trichoptera under laboratory conditions. However, due to the low level of Cry toxins in aquatic systems (Douville et al., 2005), and the rapid decomposition of maize leaves (Griffiths et al., 2009), exposure of Trichopterans in aquatic ecosystems is likely to be low in many cases.

In an acute toxicity test, Cry3Bb1 toxin from MON863 maize root extracts was fed to *Chironomus dilutus* Shobanov larvae for 10 days. A significant decrease

in *C. dilutus* survival at nominal concentrations of 30 ng/ml was found; however, no effect on growth among the surviving larvae was observed (Prihoda and Coats, 2008). No direct effects on the snail, *Cantareus asperses* Müller (Stylommatophora: Helicidae), were detected after exposure to purified Cry1Ab toxin for 4 weeks (Kramarz *et al.*, 2007a) or to growing *Bt* and non-*Bt* maize for 3 months in microcosm experiments, though the Cry1Ab toxin was detected in snail faeces and thus it represents an additional route of exposure for soil microorganisms (de Vaufleury *et al.*, 2007). In a no-choice feeding experiment at the end of growth (47 weeks of exposure), snails exposed to Cry1Ab toxin in food and soil had a growth coefficient 25 % lower than unexposed snails (Kramarz *et al.*, 2009). After the first period of reproduction (68 weeks), a significant difference remained for body mass growth between the two treatments. Differences in body mass were not significant at the end of exposure (88 weeks).

Indirect effects of GMPs on birds were studied by Gibbons *et al.* (2006) and by Chamberlain *et al.* (2007) based on data from the British farm-scale trials. The study aimed to compare bird abundance between GM herbicide-tolerant and conventional crops. The observed differences were in agreement with likely differences in food availability (Chamberlain *et al.* 2007). Moreover, using farmland birds as a model system, Butler *et al.* (2007) have developed a generic risk assessment framework that accurately predicts the current conservation status of each bird species and population growth rate associated with past changes in UK agriculture. They concluded that replacing equivalent conventional crops in the current UK agricultural landscape with GMHT crops would only have a limited effect on the Farmland Bird Index (a measure of their biodiversity). Forecasts about possible effects at landscape levels can be obtained when spatial and temporal models incorporating agricultural landscaped and their cropping patterns are available (Castellazzi *et al.*, 2007).

4. LACK OF KNOWLEDGE AND FURTHER IMPROVEMENT OF RISK ASSESSMENT PROCEDURES

Even though the scientific literature dealing with non-target organisms and GMPs is continuously increasing, there are still knowledge gaps that will need to be addressed in future studies. Table 3 lists the number of species of natural enemies and pollinators subjected so far to specific studies using GMPs and/or their products. Several new species were studied in the last few years, nevertheless there are some important groups that are still not, or only poorly, represented. In spite of their relevance in some agro-ecosystems, no information on predaceous

Diptera or Odonata are available, and only 2 spider and 1 spider mite species have been studied to date. Also, Coleoptera Staphilinidae are unfortunately lacking in the list of organisms considered so far in biosafety research. This guild constitutes, for instance, the third most abundant group of soil-dwelling arthropods in maize crops in Spain, and it has a species richness and diversity similar to ground beetles and spiders (Farinós *et al.*, 2008). Similarly, obvious knowledge gaps persist for groups of parasitoids of relevant economic importance in several agroecosystems (e.g. Eulophidae, Aphelinidae, Tachinidae) and establishing their compatibility with the use of GM crops would be very important for integrated pest management programs. When it comes to pollinators, information is only available concerning honeybees, 3 bumblebee species and the red mason bee. In some areas, solitary bees may have a very important ecological role, especially when crops are cultivated near natural habitats (Arpaia *et al.*, 2006b), therefore important information might be gathered in more specific studies.

Also when information about relevant non-target species is available, risk assessment of non-target organisms can be improved by considering their ecology in natural conditions better. Charleston and Dicke (2008) reviewed some of the dossiers for commercialisation of GMPs submitted to the Dutch National Competent Authority and highlighted several serious shortcomings in the tests conducted on non-target organisms. Firstly, the choice of assessment endpoints was often limited to the detection of acute toxic effects, and sub-lethal effects were ignored. While mortality is obviously a main life history factor to be measured, it is important to consider that sub-lethal effects alone can also drive a population to extinction (Hallam et al., 1993). Therefore, other measurement endpoints such as development, growth, fecundity, fertility, etc. need to be considered to predict any possible environmental effect. Secondly, even when the selected species for testing was considered appropriate, in several cases the life stage selected may not have been entirely appropriate (Charleston and Dicke, 2008). A sound analysis of the possible exposure in field conditions (e.g. which stage is used as a prey in the experiment, which stage of the non-target is exposed, whether bi-trophic or tri-trophic exposure is more likely, etc.) is very important in order to properly design experiments. Last, but not least, a properly designed experiment with the sufficient power to detect any adverse effect is fundamental to collect meaningful information for environmental risk assessment (Andow, 2003; Perry et al., 2009).

Table 3. Number of species of natural enemies and pollinators studied for testing the effects of genetically modified plants and/or their products. Note: species which were only surveyed in field trials as part of species assemblages are not included. The list is complete until July 2009.

Functional group	Order	Family	No. of species
Predators	Heteroptera	Anthocoridae	4
11	и	Nabidae	1
"	"	Geocoridae	2
"	н	Miridae	2
"	"	Reduvidae	1
"	"	Pentatomidae	1
"	Coleoptera	Coccinellidae	9
"	п	Carabidae	17
"	Neuroptera	Chrysopidae	1
"	Araneae	Araneidae	2
"	Acarina	Phytoseidae	1
Parasitoids	Hymenoptera	Braconidae	8
"	н	Ichneumonidae	3
"	н	Eulophydae	1
"	п	Aphelynidae	1
"	п	Encyrtidae	1
"	п	Trichogrammatidae	1
"	п	Apidae	5

Many discussions about possible negative environmental effects due to the cultivation of GMPs identified the need of predicting long-term impacts of these plants (e.g. the possible development of resistant pest strains, build up of populations of secondary pests, etc.). Long-term effects may be difficult to measure as the environment is dynamic and agro-ecosystems undergo substantial change due to varietal improvement, agronomic innovation and climatic shifts. In addition, agro-ecosystems have a relatively short time-scale over which they remain in cultivation. Some of the possible effects, e.g. linked to a chronic exposure to a particular GM plant that results in a delayed response by organisms (or their progeny), might possibly be assessed in confined experimental systems maintained over several generations. In this case the selection of appropriate measurement endpoints based on the biology of the chosen organism is essential. Other effects that may occur at a later stage due to spatial and temporal complexities are not likely to be revealed in highly confined experimental systems. In these cases, research studies together with modelling and monitoring are appropriate tools to investigate possible long-term environmental effects resulting from GMO cultivation (BVL, 2009).

5. ANALYSIS OF RISK ASSESSMENT APPROACHES

Testing of non-target organisms is deemed necessary in risk assessment in order to evaluate possible direct and indirect effects of the environmental release of GMPs. However, there are broad discussions on the potential and flexibility of different approaches and the nature of the conclusions derived from different testing regimes (see EFSA, 2008). Several risk assessment approaches have been proposed for the analysis of the possible impacts on non-target organisms; the two illustrated here represent alternative approaches to which, even with some modifications, many other risk assessment proposals refer to.

The tiered eco-toxicological approach using clearly defined risk hypotheses and the selection of surrogate species is suggested for instance by Romeis *et al.* (2008). This approach was originally developed in pesticide toxicology and is currently applied to insect resistant plants in the USA. In this approach, the potentially toxic product expressed in the GMP is considered the stressor to be characterised. The authors consider necessary to select species which are representative of their genera and/or of particular functional groups (including herbivores, pollinators, predators and parasitoids, decomposers of plant material) to serve as surrogates that can be tested under laboratory and/or field conditions and which represent ecologically and economically important animal taxa likely to be exposed to the GM crop or its products. The availability of standard laboratory tests is an important requirement, and guides surrogate species selection. As a first step ('tier'), very high doses of the toxin ("worst case" conditions) are used to test the sensitivity of surrogate species to the newly-expressed products in GMPs. Any insensitivity of tested species to high toxin doses in this framework implies that no effect will occur even under field exposure on these species, and therefore no further testing is required. By contrast, any species found to be sensitive are assessed further in more detail and in semi-field and field experiments (higher 'tiers'), in order to verify whether negative effects detected in laboratory conditions will occur in more realistic settings. Field studies are therefore not always considered necessary in this approach. In addition, as tests move from the lab to the field, it is acknowledged that more sources of potential experimental variation are introduced and thus can increase the difficulty in confirming causal relationships in the field. Based on the experience with Cry toxins, tier 1 tests appear to represent useful predictors for results at higher tier tests (Duan et al., 2010) providing that designs include all ecologically-relevant routes of exposure (e.g. including tri-trophic experiments with Bt plants). However, studies with soildwelling organisms performed in a multi-year project in the EU (Birch et al., 2007) do not seem to be in agreement with this general trend.

An "ecological approach" to the risk assessment of NTOs was originally proposed by Andow and Hilbeck (2004) and further elaborated in a series of case studies of environmental risk assessment of Bt-expressing crops in developing countries (Hilbeck and Andow, 2004; Hilbeck et al., 2006; Andow et al., 2008). The approach stresses the differences between environmental exposure of a chemical product and of a toxic compound expressed in a GMP. The approach requires that effects of the GMP be tested on a few important non-target species specifically selected from the range of environments where the GM crop is likely to be grown. Several additional criteria are considered in a selection matrix, helping to rank the importance of each candidate species in the specific agroecosystems. This risk assessment scheme further requires a process relying on risk hypotheses to guide the characterisation of exposure, adverse effects and risk, and a dynamic and adaptive tiered process where field studies are considered an essential component (Andow et al., 2006b). The obligatory integration of field studies is deemed essential for introducing ecological realism, which occurs in the more complex spatial-temporal environment because laboratory trials might not capture any potential significant environment-organism interaction. Another reason for advocating field studies is represented by the possible synergistic effect (e.g. for cry toxin) with the presence of other environmental stressors that

may enhance toxic effects (e.g. Brousseau *et al.*, 1998; Wraight and Ramos, 2005; Lawo *et al.*, 2008) and that are not normally included in laboratory studies.

While these two approaches were mostly considered mutually exclusive, it can be recognised that there are objective merits in both; for instance the growing knowledge about some specific GM events might be handled with some confidence with an eco-toxicological approach, while the ecological approach might better fulfil the Precautionary Principle (Principle 15 of the "Rio Declaration"; General Assembly of the United Nations, 1992) for new available events. As a follow up of the Scientific Colloquium on Environmental Risk Assessment of GMPs (EFSA, 2008), the European Food Safety Authority committed to produce a scientific document, currently in preparation, which incorporates some aspects from both approaches. The implications for risk assessment are manifold and a detailed discussion goes beyond the scope (and the length) of the present review. However some general recommendations can be made, in particular it may be useful to recall here two points that may be critical in properly framing a risk assessment on NTOs. It is very important to determine what type of agriculture is considered as the comparator for the specific receiving environment. The appropriate basis for comparison should be determined during the initial phase of an environmental risk assessment (USA EPA, 1998), and the current agricultural practices should be considered (Arpaia, 2004) especially referring to pest management measures. Additional factors that may influence this determination include the policy goals of the regulatory authority and the potential users of the technology (Andow et al., 2006a).

The use of mathematical models for risk assessment is widespread, however it is not trivial to remember that models are only as good as the data which drive them and thus risk assessors should ensure that dependable empirical data informs any modelling presented. This should be achieved by more cooperation between modellers and data-gatherers to ensure that expensive field trials generate data which not only help inform the environmental risk assessment but also help develop/validate models, which in turn may add confidence to risk characterisation and field trial design (EFSA, 2008).

6. FUTURE TRAITS

According to the latest available official data on the commercialisation of GM crops (James, 2009), the adoption of GM crops expressing multiple characters (stacked events) is increasing relative to other traits; a total of 28.7 million hectares

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of stacked biotech crops were planted in 2009 (up from 26.9 million hectares in 2008). Insect resistance currently represents the third diffused category of traits introduced in GMPs (Figure 1) accounting for 15 % of the cultivated GMPs worldwide.





This trend has clear implications for the risk assessment of GMPs on non-target organisms. When dealing with GMPs featuring insect resistance characteristics, the risk hypotheses considered were rather specific (e.g. a Cry toxin might not be as specific as predicted, and therefore hazards on other taxa need to be ascertained). With GMPs expressing other characteristics and producing metabolites whose toxic properties are sometimes not known, the consideration of more generic risk hypotheses (e.g. will the GMP cultivation impair predator activity?) should be included in the initial problem formulation phase. Unanticipated effects during the development of new GMPs might arise due to pleiotropy, positional effects, *in vitro* regeneration, re-arrangement in

metabolic patterns, etc. This is because transgene integration in plants occurs through illegitimate recombination. DNA integration is random, with a preference for generic regions, and gene disruptions, sequence changes and the production of new proteins can occur as a consequence of the recombination event (Rischer and Oksman-Caldentey, 2006). When such changes result in significant phenotypic modifications of the agronomic characteristics, these events will certainly be eliminated by the selection processes of breeders and companies. On the other hand, such changes may remain unnoticed if these metabolic rearrangements do not trigger unfavorable phenotypic changes.

Plant-insect interactions are largely mediated by chemical clues, even though colours, shapes, textures, etc. also play an important role. Insect herbivores and plants have co-evolved in order to optimise, each for its own benefit, the interpretation of chemical clues to successfully occupy their ecological niche. Therefore familiar food webs are ultimately constructed thanks also to the fine tuning, at all trophic levels, of such chemical perception. Under this scenario, it seems logical to hypothesise that the strict observation of an eco-toxicological approach for risk assessment on NTO will unlikely be sufficient, for instance, for GMPs in which some metabolic pathways were purposefully altered (e.g. GMPs for the production of industrial or medicinal products, energy production, phytoremediation, etc.). In the near future GMPs will likely express different metabolites for which there is much less experience in toxicological studies. In order to rule out the possibility of unintended effects linked to the genetic transformation, more general risk hypotheses will have to be considered. It has to be reflected that both the newly-produced metabolites and the GM plant itself might potentially represent an environmental stressor to be introduced in the agro-ecosystem. The author believes that in planta tests with NTOs are paramount when working with GM plants, such as pharma plants or plants with post-transcriptional gene silencing mechanisms, with which risk assessors have limited experience and need to predict their possible ecological implications. A further consideration of a comparative analysis of chemical composition of GM and control plants may also help in this assessment, since other groups of metabolites that are only partly considered during food or feed safety assessments become important in regulating plant relationships with arthropods (e.g. nutrients in all plant parts, secondary metabolites involved in plant direct defences, volatiles constituting indirect plant defences, etc.).

7. DEVELOPING COUNTRIES

Agricultural biotechnologies have the potential of meeting food and agricultural needs in developing countries. However, potential environmental

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risks and benefits need to be taken into account when making decisions about the use of GMOs, especially in highly bio-diverse countries mostly located in the tropics. For those developing countries which are also important centres of origin for the major crop species, a thorough environmental risk assessment is extremely important before the cultivation of GM plants is considered.

Lövei et al. (2009) considered the geographical distribution of the published studies dealing with NTOs in laboratory experiments. A detailed analysis of the database demonstrated that no NTO species endemic to the continents of South America. Africa or Australia have been subject to detailed laboratory studies, as compared to species populating the same agro-ecosystems in the USA, Western Europe and China. Basic biological knowledge is clearly lacking in many cases, even though the situation is far from being uniform in this respect. During two specific workshops held to plan an environmental risk assessment for the adoption of *Bt* maize in Kenya and *Bt* cotton in Brazil, several groups of non-target organisms were considered in order to start a species selection process. Pollinators and pollen feeders were identified, amongst others, as important guilds likely to be exposed to GMPs and therefore included in the analysis. In the case of Bt maize in Kenya, the working group agreed that no systematic observations on flower-visiting species associated with maize had been conducted in Kenva and they could only come up with a short list of some pollen-feeders known to be present in significant numbers in maize cultivations (Birch et al., 2004). The outcome of a similar exercise was quite different for Bt cotton in Brazil, where a working group could produce a list of several antophylous species including bumblebees, honeybees, stingless bees, solitary bees, orchid bees and several pollen feeders including ladybeetles, boll weevil, predatory mites, lacewings, earwigs (Arpaia et al., 2006b).

Fragile ecosystems tend to be characterised by high rates of species turnover and high fluctuations in population sizes (Nilsson and Grelsson, 1995); features which are often associated with agro-ecosystems in developing countries, particularly those in marginal climates for agriculture (Grant, 1989). It is generally recognised that there is a lack of baseline bio-ecological information and limited local funds and trained personnel for studies to generate such data. Moreover, specific local expertise to perform environmental risk assessment and post-release monitoring programs for GMPs is often lacking in developing countries. These needs are the focus of

several capacity building projects currently operating in these regions. There are great expectations that GM traits addressing the primary agricultural needs of developing countries (e.g. drought tolerance) will become widely available in the future. It is therefore important that the benefit-sharing objective stated in the Convention on Biological Diversity will be pursued in order to ensure that developing countries will benefit from biotechnology without endangering their natural resources and their ecosystems.

8. CONCLUDING REMARKS

The commercialisation of genetically modified plants is probably the innovation that has experienced the fastest rate of adoption ever in agriculture, in both developed and developing countries (James, 2009). However, the uneven adoption of GMP cultivation around the world reveals differences in societal attitudes toward these crops, for instance, in Europe as opposite to the USA (Marshal, 2009). The political debate repeatedly involves governments, regulators, farmers, traders as well as non-governmental organisations, environmentalists and consumers associations. Examples are reported daily by the press and media in many areas of the world.

From a researcher's perspective, it is always to be hoped that the distinction between rigorous scientific reasoning and the socio-economic interests of the various stakeholders is clearly differentiated in ongoing discussions. There are still a number of differing legitimate views and interests in the scientific community in the area of potential environmental risks linked to the use of GMOs, and an attempt has been made by the author to highlight areas of uncertainties where science could furnish further inputs to existing knowledge and to help regulators.

As already stated elsewhere (Andow *et al.*, 2006a), the author reiterates his optimism that "the rapidly accumulating base of empirical knowledge will soon make possible that the likelihood of realizing most benefits of GMPs is increased and the likelihood of environmental harm is reduced." As with any other technology, risks to the environment are possible, and in some cases practical ideas for their management can be proposed (EFSA, 2009). On the other hand, environmental benefits appear achievable with the cultivation of GMPs; for example, the reduction of insecticide use is certainly a remarkable goal that seems realistic in several conditions (e.g. Morse *et al.*, 2005; Kleter *et al.*, 2007; Naranjo 2009). Agriculture has a long record of affecting biodiversity and its functioning at several levels, and a major change, for instance, in pest management practices will certainly induce a shift (either positive or negative) on biodiversity. This review proposes an approach to a scientifically-sound consideration

of the receiving environments where GMPs are to be released; this approach can be useful in summarising relevant information, identifying knowledge gaps and set up regional environmental risk assessment programs.

Plants provide the basis for complex food webs which host hundreds of different organisms; not all of these organisms can realistically become the subject of specific studies. The analysis of the main ecosystem services provided to agriculture can represent the underlying criterion enabling the estimation of the possible environmental impacts of GMP cultivation. In every agro-ecosystem, key functions such as plant pollination, natural pest control and maintaining soil fertility are mediated by NTOs, and thus represent pillars of farming systems. It is therefore suggested that the main functional groups involved in these functions be studied for their response to GMPs at the level of organism, colony and population, and by thoroughly considering both species abundance and diversity. Anthropocentric values should also be considered. The author considers that a scientifically-sound regulatory framework which considers the ecology of the agro-ecosystems in which a GMP is to be authorised for released is the best guarantee that biotechnological innovations, as well as other future environmental challenges, can be managed in a sustainable way without renouncing their benefits.

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9. REFERENCES

Ahmad A, Gerald E, Wilde R, Whitworth J & Zolnerowich G 2006. Effect of corn hybrids expressing the coleopteran-specific Cry3Bb1 protein for corn rootworm control on aboveground insect predators. *Journal of Economic Entomology* 99(4): 1085-1095.

Álvarez-Alfageme F, **Ferry** N, **Castañera** P, **Ortego** F & **Gatehouse** AMR 2008. Prey mediated effects of Bt-maize on fitness and digestive physiology of the red spider mite predator *Stethorus punctillum* Weise (Coleoptera: Coccinellidae). *Transgenic Research* 17: 943-954.

Álvarez-Alfageme F, **Ortego** F & **Castañera** P 2009. Bt-maize fed-prey mediated effect on fitness and digestive physiology of the ground predator *Poecilus cupreus* L. (Coleoptera: Carabidae). *Journal of Insect Physiology* 55: 143-149.

Anderson PL, **Hellmich** RL, **Sumerford** DV & **Lewis** LC 2004. Effects of Cry1Abexpressing corn anthers on monarch butterfly larvae. *Environmental Entomology* 33: 1109-1115.

Andow DA 2003. Negative and positive data, statistical power, and confidence intervals. *Environmental Biosafety Research* 2: 75–80.

Andow DA & **Hilbeck** A 2004. Science-based risk assessment for non target effects of transgenic crops. *Bioscience* 54: 637–649.

Andow DA, **Lövei** GL & **Arpaia** S 2006a. Ecological risk assessment for Bt crops. *Nature Biotechnology* 24: 749-751.

Andow DA, Birch NAE, Dusi AN, Fontes EMG, Hilbeck A, Lang A, Lövei GL, Pires CSS, Sujii ER, Underwood E & Wheatley RE 2006b. Non-target and biodiversity ecological risk assessment for GM crops. In: *Proceedings of* 9th International Symposium on the Biosafety of Genetically Modified Organisms, International Society for Biosafety Research, September 24-29 2006, Jeju Island, Korea, pp 68-73.

56

Andow DA, **Hilbeck** A & **Van Tuàt** N 2008. Environmental Risk Assessment of Genetically Modified Organisms: Vol. 4. Challenges and opportunities with Bt Cotton in Vietnam, CABI Publishing, Wallingford, UK.

Arpaia S 1996. Ecological impact of Bt-transgenic plants: 1. Assessing possible effects of CryIIIB toxin on honey bees. *Journal of Genetics and Breeding* 50: 315-319.

Arpaia S 2004. Workshop report - Monitoring/Bioindicators. *IOBC-WPRS Bullettin* 27(3): 205-208.

Arpaia S, **Di Leo** GM, **Guerrieri** E & **Fiore** MC 2006a. Parasitization of *Chromatomia horticola* Goreau in experimental fields with genetically modified canola. *IOBC-WPRS Bullettin* 29(5): 21-26.

Arpaia S. **Fonseca** VLI, **Pires** CS & **Silveira** FA 2006b. Non-Target and Biodiversity Impacts on Pollinators and Flower-Visiting Insects. In: *Environmental Risk Assessment of Genetically Modified Organisms: Vol. 2. Methodologies for Assessing Bt Cotton in Brazil.* Hilbeck A, Andow DA & Fontes EMG (eds.), CABI Publishing, Wallingford, UK. Pp 155-174.

Arpaia S, **Di Leo** GM, **Fiore** MC, **Schmidt** JEU & **Scardi** M 2007. Composition of arthropod species assemblages in Bt-expressing and near isogenic eggplants in experimental fields. *Environmental Entomology* 36(1): 213-227.

Arpaia S, **Schmidt** JEU, **Di Leo** GM & **Fiore** MC 2009. Oviposition of the Colorado potato beetle (*Leptinotarsa decemlineata*) and natural predation on its egg masses in Bt-expressing fields. *Biocontrol Science and Technology* 19(9): 971-984.

Babendreier D, **Kalberer** NM, **Romeis** J, **Fluri** P & **Bigler** F 2004. Pollen consumption in honey bee larvae: a step forward in the risk assessment of transgenic plants. *Apidologie* 35: 293-300.

Babendreier D, **Kalberer** NM, **Romeis** J, **Fluri** P, **Mulligan** E & **Bigler** F 2005. Influence of Bt-transgenic pollen, Bt-toxin and protease inhibitor (SBTI) ingestion on development of the hypopharyngeal glands in honeybees. *Apidologie* 36: 585-594.

Bai YY, **Jiang** MG & **Cheng** JA 2005. Effects of transgenic cry1Ab rice pollen on fitness of *Propylea japonica* (Thunberg). *Journal of Pesticide Science* 78: 123-128.

Bakonyi G, **Szira** F, **Kiss** I, **Villányi** I, **Seres** A & **Székács** A 2006. Preference tests with collembolas on isogenic and Bt-maize. *European Journal of Soil Biology* 42: 132-135.

Baumgarte S & **Tebbe** CC 2005. Field studies on the environmental fate of the Cry1Ab Bt-toxin produced by transgenic maize (MON810) and its effect on bacterial communities in the maize rhizosphere. *Molecular Ecology* 14: 2539-2551.

Birch ANE, Wheatley R, Hilbeck A, Arpaia S, Lövei G, Songa J, Sétamou M, Sithanantham S, Ngi-Song A, Anyango B, Muyekho F, Ochieno D, Lelmen E, Kalama P, Degaga E, Soares Melo I, Pitelli R, Sujii E, Capalbo D, Fontes EGM, Wan F, Ogwang J, Smith J, Van Son N & Quang Tan T 2004. Biodiversity and Non-Target Impacts: a Case Study of Bt-maize in Kenya. In: *Environmental Risk Assessment of Genetically Modified Organisms, Volume 1: A Case Study of Bt-maize in Kenya.* Hilbeck A & Andow D [eds.]. CABI Publishing, Wallingford, UK. pp. 117-185.

Birch ANE, **Griffiths** BS, **Caula** S, **Thompson** J, **Heckmannb** LH, **Krogh** PH & **Cortet** J 2007. The role of laboratory, glasshouse and field scale experiments in understanding the interactions between genetically modified crops and soil ecosystems: A review of the ECOGEN project. *Pedobiologia* 51: 251–260.

Bøhn T, **Primicerio** R, **Hessen** D & **Traavik** T 2008. Reduced fitness of Daphnia magna fed a Bt-transgenic maize variety. Archives of Environmental Contamination and Toxicology 55: 584-592.

Bourguet D, **Chaufaux** J, **Micoud** A, **Delos** M, **Naibo** B, **Bombarde** F, **Marque** G, **Eychenne** N & **Pagliari C** 2002. Ostrinia nubilalis parasitism and the field abundance of non-target insects in transgenic *Bacillus thuringiensis* corn (*Zea mays*). Environmental Biosafety Research 1: 49–60.

Brooks DR, Bohan DA, Champion GT, Haughton AJ, Hawes C, Heard MS, Clark SJ, Dewar AM, Firbank LJ, Perry JN, Rothery P, Scott RJ, Woiwod IP, Birchall C, Skellern MP, Walker JH, Baker P, Bell D, Browne EL, Dewar ALG, Fairfax CM, Garner BH, Haylock LA, Horne SL, Hulmes SE, Mason NS, Norton LR, Nuttall P, Randle Z, Rossall MJ, Sands RJN, Singer EJ & Walker MJ 2003. Invertebrate responses to the management of genetically modified herbicide -tolerant and conventional spring crops. I. Soil-surface-active invertebrates. *Philosophical Transactions of the Royal Society London B* 358: 1847-1862.

Brousseau C, **Charpentier** G & **Belloncik** S 1998. Effects of *Bacillus thuringiensis* and destruxins (*Metarhizium anisopliae* mycotoxins) combinations on spruce budworm (Lepidoptera: Tortricidae). *Journal of Invertebrate Pathology* 72: 262–268.

Brussaard L, **de Ruiter** PC & **Brown** GG 2007. Soil biodiversity for agricultural sustainability. *Agriculture, Ecosystems and Environment* 121: 233–244.

Burgess EPJ, **Malone** LA & **Christeller** JT 1996. Effects of two proteinase inhibitors on the digestive enxymes and survival of honey bees (*Apis mellifera*). Journal of Insect Physiology 42: 823-828.

Burgio G, Lanzoni A, Accinelli G, Dinelli G, Bonetti A, Marotti I & Ramilli F 2007. Evaluation of Bt-toxin uptake by the non-target herbivore, *Myzus persicae* (Hemiptera: Aphididae), feeding on transgenic oilseed rape. *Bulletin of Entomological Research* 97: 211–215.

Burns RC & **Dick** RP 2002. Enzymes in the environment: Activity, ecology, and applications. Marcel Dekker, NY. **Butler** SJ, Vickery JA & Norris K 2007. Farmland biodiversity and the footprint of agriculture. *Science* 315(5810): 381 – 384.

BVL 2009. BEETLE report. Long-term effects of genetically modified (GM) crops on health and the environment (including biodiversity): Prioritization of potential risks and delimitation of uncertainties. German Federal Office of Consumer Protection and Food Safety (BVL), BLaU-Umweltstudien and Genius GmbH, 133 pp. Available at http://ec.europa.eu/environment/biotechnology/pdf/beetle_report.pdf. Accessed online March 2010.

Castaldini M, Turrini A, Sbrana C, Benedetti A, Marchionni M, Mocali S, Fagiani A, Landi S, Santomassimo F, Pietrangeli B, Nuti MP, Miclaus N & Giovanetti M 2005. Impact of Bt corn on rhizospheric and soil eubacterial communities and on beneficial mycorrhizal symbiosis in experimental microcosms. *Applied and Environmental Microbiology* 71: 6719-6729.

Castellazzi MS, **Perry** JN, **Colbach** N, **Monod** H, **Adamczyk** K, **Viaud** V & **Conrad** KF 2007. New measures and tests of temporal and spatial pattern of crops in agricultural landscapes. *Agriculture, Ecosystems and Environment* 118: 339–349.

Chamberlain DE, **Freeman** SN & **Vickery** JA 2007. The effects of GMHT crops on bird abundance in arable fields in the UK. *Agriculture, Ecosystems and Environment* 118: 350–356.

Chambers CP, **Whiles** MR, **Griffiths** NA, **Evans-White** MA, **Rosi-Marshall** EJ, **Tank** JL & **Royer** TV 2007. Assessing the impacts of transgenic Bt corn detritus on macroinvertebrate communities in agricultural streams. In: *Proceedings of North American Benthological Society 55th Annual Meeting*, June 3-8, 2007 Columbia, SC, USA, pp. 373.

Charleston DS & **Dicke** M 2008. Designing experimental protocols to investigate the impact of GM crops on non-target arthropods. COGEM onderzoeksrapport CGM 2008-01. Available at http://www.cogem.net/ContentFiles/CGM%202008-01%20Niet%20doelwit%20organismen.pdf.

60

Accessed online March 2010.

Clark BW & **Coats** JR 2006. Subacute effects of Cry1Ab Bt corn litter on the earthworm *Eisenia foetida* and the springtail *Folsomia candida*. *Environmental Entomology* 35: 1121–1129.

Clark BW, **Prihoda** KR & **Coats** JR 2006. Subacute effects of transgenic Cry1Ab Bacillus thuringiensis corn litter on the isopods Trachelipus rathkii and Armadillidium nasatum. Environmental Toxicology and Chemistry 25: 2653-2661.

Cloutier C, **Boudreault** S & **Michaud** D 2008. Impact de pommes de terre résistantes au doryphore sur les arthropodes non visés: une méta-analyse des facteurs possiblement en cause dans l'échec d'une plante transgénique Bt. *Cahiers Agricultures* 17(4): 388-394.

Cortet J, Andersen MN, Caul S, Griffiths B, Joffre R, Lacroix B, Sausse C, **Thompson** T, Krogh PH (2006). Decomposition processes under Bt (*Bacillus thuringiensis*) maize: Results of a multisite experiment. Soil Biology & Biochemistry 38: 195–199.

Costanza R, **D'arge** R, **De Groot** R, **Farber** S, **Grasso** M, **Hannon** B, **Limburg** K, **Naeem** S, **O'neill** RV, **Paruelo** J, **Raskin** RG, **Sutton** P **& Van Den Belt** M 1997. The value of the world's ecosystem services and natural capital. *Nature* 387: 253-260.

Cowgill SE, & **Atkinson** HJ 2003. A sequential approach to risk assessment of transgenic plants expressing protease inhibitors: effects on nontarget herbivorous insects. *Transgenic Research* 12: 439-449.

Crane E & **Walker** P 1986. Pollination directory for world crops. International Bee Association. *New Zealand Journal of Botany* 24: 355-356.

Curtis TP, **Sloan** WT & **Scannell** JW 2002. Estimating prokaryotic diversity and its limits. *Proceedings of the National Academy of Science USA* 99: 10494-10499.

Daily GC 1997. Nature's services. Island Press, Washington, DC, USA.

Debach P & **Rosen** D 1991. *Biological Control by Natural Enemies*. Second edition. Cambridge University Press, UK.

de la Poza M, Pons X, Farinós GP, López C, Ortego F, Eizaguirre M, Castañera P & Albajes R 2005. Impact of farm-scale Bt-maize on abundance of predatory arthropods in Spain. *Crop Protection* 24: 677-684.

de Vaufleury A, **Kramarz** PE, **Binet** P, **Cortet** J, **Caul** S, **Andersen** MN, **Plumey** E, **Coeurdassier** M & **Krogh** PH 2007. Exposure and effect assessments of *Bt*-maize on non-target organisms (gastropods, microarthropods, mycorrhizal fungi) in microcosms. *Pedobiologia* 51: 185-194.

Dhillon MK & **Sharma** HC 2009. Effects of *Bacullus thuringiensis* -endotoxins Cry1Ab and Cry1Ac on the coccinellid beetle, *Cheilomenes sexmaculatus* (Coleoptera, Coccinellidae) under direct and indirect exposure conditions. *Biocontrol Science and Technology* 19: 407-420.

Dively GP, **Rose** R, **Sears** MK, **Hellmich** RL, **Stanley-Horn** DE, **Calvin** DD, **Russo** JM & **Anderson** PL 2004. Effects on monarch butterfly larvae (Lepidoptera: Danaidae) after continuous exposure to Cry1Ab-expressing corn during anthesis. *Environmental Entomology* 33: 1116-1125.

Douville M, **Gagné** F, **Masson** L, **McKay** J & **Blaise** C 2005. Tracking the source of *Bacillus thuringiensis* Cry1Ab endotoxin in the environment. *Biochemical Systematics and Ecology* 33: 219-232.

Duan JJ, **Marvier** M, **Huesing** J, **Dively** G & **Huang** ZY 2008. A meta-analysis of effects of Bt crops on honey bees (Hymenoptera: Apidae). *PLoS ONE* 3: 1-6 (e1415).

Duan JJ, **Lundgren** JG, **Naranjo** S & **Marvier** M 2010. Extrapolating nontarget risk of Bt crops from laboratory to field. *Biology Letters* 6: 74–77. **Duelli** P & **Obrist** MK 1998. In search of the best correlates for local organismal biodiversity in cultivated areas. *Biodiversity and Conservation* 7: 297-309.

Dutton A, **Klein** H, **Romeis** J & **Bigler** F 2002. Uptake of Bt-toxin by herbivores feeding on transgenic maize and consequences for the predator *Chrysoperla carnea. Ecological Entomology* 27: 441- 447.

Dutton A, **Romeis** J & **Bigler** F 2003. Assessing the risks of insect resistant transgenic plants on entomophagous arthropods: Bt-maize expressing Cry1Ab as a case study. *BioControl* 48: 611 - 636.

Eckert J, **Schuphan** I, **Hothorn** LA & **Gathmann** A 2006. Arthropods on maize ears for detecting impacts of Bt-maize on nontarget organisms. *Environmental Entomology* 35: 554-560.

Escher N, **Kach** B & **Nentwig** W 2000. Decomposition of transgenic *Bacillus thuringiensis* maize by microorganisms and woodlice *Porcello scaber* (Crustacea: Isopoda). *Basic and Applied Entomology* 1: 161-169.

EFSA 2008. Environmental risk assessment of Genetically Modified Plants – Challenges and approaches. EFSA Scientific Colloquium Summary Report, European Food Safety Authority (EFSA), Parma, Italy. Available at http://www.efsa.europa.eu/en/colloquiagmoera/publication/comm_ colloquium8_2008_en.pdf. Accessed online March 2010.

EFSA 2009. Scientific Opinion of the Panel on Genetically Modified Organisms on Applications (EFSA-GMO-RX-MON810) for renewal of authorisation for the continued marketing of (1) existing food and food ingredients produced from genetically modified insect resistant maize MON810; (2) feed consisting of and/or containing maize MON810, including the use of seed for cultivation; and of (3) food and feed additives, and feed materials produced from maize MON810, all under Regulation (EC) No 1829/2003 from Monsanto. *The EFSA Journal* 1149: 1-85.

Fang M., **Kremer** RJ, **Motavalli** PP & **Davis** G 2005. Bacterial diversity in rhizosphere of nontransgenic and transgenic corn. *Applied Environmental Microbiology* 71: 4132-4136.

Fang M, Motavalli PP, Kremer RJ & Nelson KA 2007. Assessing changes in soil microbial communities and carbon mineralieation in Bt and non-Bt corn residue-amended soils. *Applied Soil Ecology* 37: 150-160.

Faria CA, Wackers FL, Pritchard J, Barrett DA & Turlings TCJ (2007). High Susceptibility of Bt-maize to Aphids Enhances the Performance of Parasitoids of Lepidopteran Pests. PLoS ONE 2(7) e600 DOI:10.1371.

Farinós GP, **de la Poza** M, **Hernandez-Crespo** P, **Ortego** F & **Castañera** P 2008. Diversity and seasonal phenology of aboveground arthropods in conventional and transgenic maize crops in Central Spain. *Biological Control* 44(3): 362-371.

Felke M, **Lorenz** N & **Langenbruch** GA 2002. Laboratory studies on the effects of pollen from Bt-maize on larvae of some butterfly species. *Journal of Applied Entomology* 126: 320-325.

Filion M 2008. Do transgenic plants affect rhizobacteria populations? *Microbial Biotechnology* 1: 463-475.

Free JB 1993. Insect Pollination of Crops. 2nd Edition. Academic Press, London, UK.

General Assembly of the United Nations 1992. Report of the United Nations Conference on Environment and Development, A/CONF.151/26 (Vol. I), 3-14 June 1992, Rio de Janeiro, Brazil. Available at http://www.un.org/documents/ ga/conf151/aconf15126-1annex1.htm. Accessed online March 2010.

Gibbons DW, Bohan DA, Rothery P, Stuart RC, Haughton AJ, Scott RJ, Wilson DJ, Perry JN, Clark SJ, Dawson RJG & Firbank LG 2006. Weed seed resources for birds in fields with contrasting conventional and genetically modified herbicide-

64

tolerant crops. Proceedings of the Royal Society B 273(1596): 1921-1928.

Garcia-Robles I, Sanchez J, Gruppe A, Martinez-Ramirez AC, Rausell C, Real MD & Bravo A (2001). Mode of action of *Bacillus thuringiensis* PS86Q3 strain in hymenopteran forest pests. *Insect Biochemistry and Molecular Biology* 31: 849–856.

Godfray HCJ 1994. *Parasitoids: Behavioral and Evolutionary Ecology*. Princeton University Press, USA.

Grant IF 1989. Monitoring insecticide side-effects in large-scale treatment programmes: tsetse spraying in Africa. In: *Pesticides and Non-Target Invertebrates*. Jepson PC (ed), Intercept, Wimborne, UK. pp. 43–71.

Griffiths BS, Caul S, Thompson J, Birch ANE, Scrimgeour C, Andersen, MN, Cortet J, Messéan A, Sausse C, Lacroix B, & Krogh PH 2005. A comparison of soil microbial community structure, protozoa, and nematodes in field plots of conventional and genetically modified maize expressing the *Bacillus thuringiensis* Cry1Ab toxin. *Plant and Soil* 275: 135-146.

Griffiths, BS, **Caul** S, **Thompson** J, **Birch** ANE, **Scrimgeour** C, **Cortet** J, **Foggo** A, **Hackett** CA & **Krogh** PH 2006. Soil microbial and faunal community responses to Bt-maize and insecticide in two soils. *Journal of Environmental Quality* 35: 734-741.

Griffiths BS, **Caul** S, **Thompson** J, **Birch** ANE, **Cortet** J, **Andersen** MN & **Krogh** PH 2007a. Microbial and microfaunal community structure in cropping systems with genetically modified plants. *Pedobiologia* 51: 195-206.

Griffiths BS, **Heckmann** LH, **Caul** S, **Thompson** J, **Scrimgeour** C, & **Krogh** PH 2007b. Varietal effects of eight paired lines of transgenic Bt-maize and nearisogenic non-Bt-maize on soil microbial and nematode community structure. *Plant Biotechnology Journal* 5: 60-68.

Griffiths NA, Tank JL, Royer TV, Rosi-Marshall EJ, Whiles MR, Chambers CP,

Frauendorf TC & **Evans-White** MA 2009. Rapid decomposition of maize detritus in agricultural headwater streams. *Ecological Applications* 19(1): 133–142.

Gupta VVSR & **Yeates** GW 1997. Soil microfauna as bioindicators of soil health. In: *Biological Indicators of Soil Health.* Pankurst CE (ed.). CAB International, New York, USA. pp. 201–233.

Hallam TG, Canziani GA & Lassiter RR 1993. Sublethal narcosis and population persistence: a modelling study on growth effects. *Environmental Toxicology and Chemistry* 12: 947-954.

Harwood JD, **Samson** RA & **Obrycki** JJ 2007. Temporal detection of Cry1Abendotoxins in coccinellid predators from fields of *Bacillus thuringiensis*. *Bulletin of Entomological Research* 97: 643-648.

Harwood JD, **Wallin** WG & **Obrycki** JJ 2005. Uptake of Bt endotoxins by non target herbivores and higher order arthropod predators: molecular evidence from a transgenic corn agroecosystem. *Molecular Ecology* 14: 2815-2823.

Haughton AJ, Champion GT, Hawes C, Heard MS, Brooks DR, Bohan DA, Clark SJ, Dewar AM, Firbank LG, Osborne JL, Perry JN, Rothery P Roy DB, Scott RJ, Woiwod IP, Birchall C, Skellern MP, Walker JH, Baker P, Browne EL, Dewar AJG, Garner BH, Haylock LA, Horne SL, Mason NS, Sands RJN & Walker MJ 2003. Invertebrate responses to the management of genetically modified herbicide -tolerant and conventional spring crops. II. Within-field epigeal and aerial arthropods. *Philosophical Transactions of the Royal Society London B* 358: 1863-1877.

Head G, **Brown** CR, **Groth** ME & **Duan** JJ 2001. Cry1Ab toxin levels in phytophagous insects feeding on transgenic corn: implications for secondary exposure risk assessment. *Entomologia Experimentalis et Applicata* 99: 37-45.

Heckmann LH, Griffiths BS, Caul S, Thompson J, Pusztai-Carey M, Moar WJ,

Andersen MN & **Krogh** PH 2006. Consequences for *Protaphorura armata* (Collembola: Onychiuridae) following exposure to genetically modified *Bacillus thuringiensis* (Bt) maize and non-Bt-maize. *Environmental Pollution* 142: 212-216.

Hilbeck A, **Baumgartner** M, **Fried** PM & **Bigler** F 1998a. Effects of transgenic Bacillus thuringiensis corn-fed prey on mortality and development time of immature Chrysoperla carnea (Neuroptera: Chrysopidae). Environmental Entomology 27: 480-487.

Hilbeck A, **Moar** WJ, **Pusztai-Carey** M, **Filippini** A & **Bigler** F 1998b. Toxicity of *Bacillus thuringiensis* Cry1Ab toxin to the predator *Chrysoperla carnea* (Neuroptera: Chrysopidae). *Environmental Entomology* 27: 1255-1263.

Hilbeck A, **Moar** WJ, **Pusztai-Carey** M, **Filippini** A & **Bigler** F 1999. Preymediated effects of Cry1Ab toxin and protoxin and Cry2A protoxin on the predator *Chrysoperla carnea*. *Entomologia Experimentalis et Applicata* 91: 305-316.

Hilbeck A & **Andow** DA 2004. Environmental Risk Assessment of Genetically Modified Organisms, Volume 1: A Case Study of Bt-maize in Kenya. CABI Publishing, Wallingford, UK.

Hilbeck A, **Andow** DA & **Fontes** EMG 2006. Environmental Risk Assessment of Genetically Modified Organisms: Vol. 2. Methodologies for Assessing Bt Cotton in Brazil. CABI Publishing, Wallingford, UK.

Hönemann L & **Nentwig** W 2009. Are survival and reproduction of *Enchytraeus albidus* (Annelida: Enchytraeidae) at risk by feeding on *Bt*-maize litter? *European Journal of Soil Biology* 45(4): 351-355.

Hopkins SP 2006. Collembola. In: *Encyclopedia of Soil Science*, Vol 1, R Lal (ed.), Taylor & Francis, UK. pp. 299-302.

Höss S, Arndt M, Baumgarte S, Tebbe CC, Nguyten HT & Jehle JA 2008. Effects of transgenic corn and Cry1Ab toxin on the nematode, *Caenorhabditis*

elegans. Ecotoxicology and Environmental Safety 70: 334-340.

Icoz I & **Stotzky** G 2008. Fate and effects of insect-resistant Bt crops in soil ecosystems. *Soil Biology & Biochemistry* 40: 559-586.

James C 2009. Global Status of Commercialized Biotech/GM Crops: 2009. ISAAA Brief No. 39. Ithaca, NY, USA.

Jasinski JR, Eisley JB, Young CE, Kovach J & Wilson H 2003. Select nontarget arthropod abundance in transgenic and nontransgenic field crops in Ohio. *Environmental Entomology* 32: 407-413.

Jesse LCH & **Obrycki** JJ 2003. Occurrence of *Danaus plexippus* L. (Lepidoptera: Danaidae) on milkweeds (*Asclepias syriaca*) in transgenic Bt corn agroecosystems. *Agriculture, Ecosystems & Environment* 97: 225-233.

Kalushkov P & **Hodek** I 2005. The effects of six species of aphids on some life history parameters of the ladybird *Propylea quatuordecimpunctata* (Coleoptera : Coccinellidae). *European Journal of Entomology* 102: 449-452.

Klein AM, Vaissière BE, Cane JH, Steffan-Dewenter I, Cunningham, SA, Kremen C & Tscharntke T 2007. Importance of pollinators in changing landscapes for world crops. *Proceedings of the Royal Society* B 274: 303-313.

Kleter GA, Bhula R, Bodnaruk K, Carazo E, Felsot AS, Harris CA, Katayama A, Kuiper HA, Racke KD, Rubin B, Shevah Y, Stephenson GR, Tanaka K, Unsworth J, Wauchope RD & Wong SS 2007. Altered pesticide use on transgenic crops and the associated general impact from an environmental perspective. *Pest Management Science* 63: 1107–1115.

Knox OGG, **Nehl** DB, **Mor** T, **Roberts** GN & **Gupta** VVSR 2008. Genetically modified cotton has no effect on arbuscular mycorrhizal colonisation of roots. *Field Crop Research* 109(1-3): 57-60.

Koziel MG, Beland GL, Bowman C, Carozzi NB, Crenshaw R, Crossland L, Dawson J, Desai N, Hill M, Kadwell S, Launis K, Lewis K, Maddox D, Mcpherson K, Meghji MR, Merlin E, Rhodes R, Warren GW, Wright M & Evola SV 1993. Field performance of elite transgenic maize plants expressing an insecticidal protein derived from *Bacillus thuringiensis*. *Bio/Technology* 11: 194 – 200.

Kramarz PE, **de Vaufleury** A & **Carey** M 2007a. Studying the effect of exposure of the snail *Helix aspersa* to the purified *Bt* toxin, Cry1Ab. *Applied Soil Ecology* 37: 169-172.

Kramarz PE, de Vaufleury A, Gimbert F, Cortet J, Tabone E, Andersen MN & Krogh PH 2009. Effects of Bt-maize material on the life cycle of the land snail *Cantareus aspersus*. *Applied Soil Ecology* 42(3): 236-242.

Krogh PH, Griffiths BS, Demšar D, Bohanec M, Debeljak M, Andersen MN, Sausse C, Birch ANE, Caul S, Holmstrup M, Heckmann LH & Cortet J 2007. Responses by earthworms to reduced tillage in herbicide tolerant maize and *Bt-maize* cropping systems. *Pedobiologia* 51: 219-227.

Lang A & **Vojtech** E 2006. The effects of pollen consumption on transgenic Bt-maize on the common swallowtail, *Papilio machaon* L. (Lepidoptera, Papilionidae). *Basic and Applied Ecology* 7: 296-306.

Lang A, Arndt M, Beck R, Bauchhenss J & Pommer G 2006. Monitoring of the environmental effects of the Bt gene. Bavarian State Research Center for Agriculture, Germany, No. 2006/10. Available at http://www.lfl-neu.bayern. de/publikationen/daten/schriftenreihe_url_1_43.pdf. Accessed online March 2010.

Lawo NCR, Mahon J, Milner RJ, Sarmah BK, Higgins TJV & Romeis J 2008. Effectiveness of *Bacillus thuringiensis*-transgenic chickpeas and the entomopathogenic fungus *Metarhizium anisopliae* in controlling *Helicoverpa armigera* (Lepidoptera: Noctuidae). Applied Environmental *Microbiology* 74:4381 – 4389.

Lilley AK, **Bailey** MJ, **Cartwright** C, **Turner** SL & **Hirsch** PR 2006. Life in earth: the impact of GM plants on soil ecology? *Trends in Biotechnology* 24: 9-14.

Losey JE, **Rayor** LS & **Carter** ME 1999. Transgenic pollen harms monarch larvae. *Nature* 399:214.

Lövei GL & **Arpaia** S 2005. The impact of transgenic plants on natural enemies a critical review of laboratory studies. *Entomologia Experimentalis et Applicata* 114: 1 – 14.

Lövei GL, **Andow** DA & **Arpaia** S 2009. Transgenic insecticidal crops and natural enemies: a detailed review of laboratory studies. *Environmental Entomology* 38(2): 293-306.

Lu BR 2008. Transgene escape from GM crops and potential biodiversity consequences: an environmental perspective. *Collection of Biosafety Reviews* 4: 66-141. Available at http://www.icgeb.org/~bsafesrv/pdffiles/ Bao-Rong.pdf. Accessed online March 2010.

Lundgren JG & **Wiedenmann** RN 2005. Tritrophic interactions among Bt (Cry3Bb1) corn, aphid prey, and the predator *Coleomegilla maculata* (Coleoptera: Coccinellidae). *Environmental Entomology* 34: 1621-1625.

Lynch J 1994. The rhizosphere – form and function. *Applied Soil Ecology* 1: 193 – 198.

Malone LA, & **Burgess** EPJ 2009. Impact of genetically modified crops on pollinators. In: *Environmental Impact of Genetically Modified Crops*, Ferry N & Gatehouse AMR (eds.). CAB International, UK. pp. 199-222.

Malone LA, **Gatehouse** AMR & **Barrat** BIP 2008. Beyond Bt: Alternative strategies for insect resistant genetically modified crops. In: *Integration of Insect-Resistant Genetically Modified Crops within IPM Programs*, Romeis J, Shelton AM & Kennedy GG (eds). Springer Science, The Netherlands. pp. 357 – 417.

70

Malone LA, **Burgess** EPJ, **Stefanovic** D, & **Gatehouse** HS 2000. Effects of four proteinase inhibitors on the survival of worker bumblebees *Bombus terrestris* L. *Apidologie* 31: 25-38.

Manachini B, & **Lozzia** GC 2002. First investigations into the effects of Bt corn crop on Nematofauna. *Bollettino di Zoologia Agraria e Bachicoltura* Serie II 34: 85–96.

Manachini B, **Landi** S, **Fiore** MC, **Festa** M & **Arpaia** S 2004. First investigations on the effects of Bt-transgenic *Brassica napus* L. on the trophic structure of the nematofauna. *IOBC/WPRS Bullettin* 27: 103–108.

Marshall A 2009. 13.3 million farmers cultivate GM crops. *Nature* biotechnology 27(3): 221.

Marvier M, **McCreedy** C, **Regetz** J & **Kareiva** P 2007. A meta-analysis of effects of Bt cotton and maize on nontarget invertebrates. *Science* 316: 1475-1477.

Mattila HR, **Sears** MK & **Duan** JJ 2005. Response of *Danaus plexippus* to pollen of two new Bt corn events via laboratory bioassay. *Entomologia Experimentalis et Applicata* 116: 31-41.

Meissle M, **Vojtech** E & **Poppy** GM 2005. Effects of Bt-maize-fed prey on the generalist predator *Poecilus cupreus* L. (Coleoptera: Carabidae). *Transgenic Research* 14: 123-132.

Mendelson M, **Kough** J, **Vaituzis** Z & **Matthews** K 2003. Are Bt crops safe? *Nature Biotechnology* 21(9): 1003–1009.

MA 2005. Ecosystems and Human Well-being: General Synthesis. Millennium Ecosystem Assessment (MA), Island Press, Washington, DC, USA.

Moonen AC & **Barberi** P 2008 Functional biodiversity: An agroecosystem approach. Agriculture, Ecosystems and Environment 127: 7–21.

Moore JC, **Walter** DE & **Hunt** HW 1988. Arthropod regulation of microand mesobiota in below-ground detrital food webs. *Annual Review of Entomology* 33: 419–439.

Morse S, **Bennett** RM & **Ismael** Y 2005. Genetically modified insect resistance in cotton: Some farm level economic impacts in India. *Crop Protection* 24: 433-440.

Naranjo SE 2005a. Long-term assessment of the effects of transgenic Bt cotton on the abundance of nontarget arthropod natural enemies. *Environmental Entomology* 34: 1193-1210.

Naranjo SE 2005b. Long-term assessment of the effects of transgenic Bt cotton on the function of the natural enemy community. *Environmental Entomology* 34: 1211-1223.

Naranjo SE 2009. Impacts of Bt crops on non-target invertebrates and insecticide use patterns. *CAB Reviews: Perspectives in Agriculture, Veterinary Science, Nutrition and Natural Resources* 4: 1-23.

Neher AD 2001. Role of nematodes in soil health and their use as indicators. *Journal of Nematology* 33(4): 161–168.

Nguyen HT & **Jehle** JA 2007. Quantitative analysis of the seasonal and tissue-specific expression of Cry1Ab in transgenic maize Mon810. *Journal of Plant Diseases and Protection* 114: 82-87.

Nilsson C & **Grelsson** G 1995. The fragility of ecosystems: A review. *Journal* of Applied Ecology 32: 677-692.

Oberhauser KS, **Prysby** M, **Mattila** HR, **Stanley-Horn** DE, **Sears** MK, **Dively** GP, **Olson** E, **Pleasants** JM, **Lam** WKF & **Hellmich** RL 2001. Temporal and spatial overlap between monarch larvae and corn pollen. *Proceedings of the National Academy of Sciences USA* 98: 11913-11918.

Obrist LB, Dutton A, Albajes R & Bigler F 2006a. Exposure of arthropod

predators to Cry1Ab toxin in Bt-maize fields. *Ecological Entomology* 31: 143-154.

Obrist LB, **Dutton** A, **Romeis** J & **Bigler** F 2006b. Biological activity of Cry1Ab toxin expressed by Bt-maize following ingestion by herbivorous arthropods and exposure of the predator *Chrysoperla carnea*. *BioControl* 51: 31-48.

O'Callaghan M, **Glare** TR, **Burgess** EPJ & **Malone** LA 2005. Effects of plants genetically modified for insect resistance on nontarget organisms. *Annual Review of Entomology* 50: 271–292.

Perry JN, Ter Braak C, Dixon PM, Duan JJ, Hails RS, Huesken A, Lavielle M, Marvier M, Scardi M, Schmidt K, Tothmeresz B, Schaarschmidt F & Van Der Voet H 2009. Statistical aspects of environmental risk assessment of GM plants for effects on non-target organisms. *Environmental Biosafety Research* 8: 65-78.

Perry JN Devos Y, Arpaia S, Bartsch D, Gathmann A, Hails RS, Kiss J, Lheureux K, Manachini B, Mestdagh S, Neemann G, Ortego F, Schiemann J & Sweet JB 2010. A mathematical model of exposure of nontarget Lepidoptera to Btmaize pollen expressing Cry1Ab within Europe. *Proceedings of the Royal Society B* 277: 1417-1425.

Peterson JA; **Obrycki** JJ; & **Harwood** JD 2009. Quantification of Bt-endotoxin exposure pathways in carabid food webs across multiple transgenic events. *Biocontrol Science and Technology* 19(6): 613-625.

Pierre J, **Marsault** D, **Genecque** E, **Renard** M, **Champolivier** J & **Pham-Delègue** MH 2003. Effects of herbicide-tolerant transgenic oilseed rape genotypes on honey bees and other pollinating insects under field conditions. *Entomologia Experimentalis et Applicata* 108: 159–168.

Pilcher CP, **Obrycki** JJ, **Rice** ME & **Lewis** LC 1997. Preimaginal development, survival and field abundance of insect predators on transgenic *Bacillus thuringiensis* corn. *Environmental Entomology* 26: 446-454.

Pleasants JM, Hellmich RL, Dively GP, Sears MK, Stanley-Horn DE, Mattila HR, Foster JE, Clark TL & Jones GD 2001. Corn pollen deposition on milkweeds in or near corn field. *Proceedings of the National Academy of Sciences* USA 98: 11919-11924.

Pont B & **Nentwig** W 2005. Quantification of Bt-protein digestion and excretion by the primary decomposer *Porcellio scaber*, fed with two Bt corn varieties. *Biocontrol Science and Technology* 15: 341-352.

Potenza C, **Aleman** L & **Sengupta-Gopalan** C 2004. Targeting transgene expression in research, agricultural, and environmental applications: Promoters used in plant transformation. *In Vitro Cell Developmental Biology - Plant* 40: 1–22.

Prasifka PL, **Hellmich** RL, **Prasifka** JR & **Lewis** LC 2007. Effects of Cry1Abexpressing corn anthers on the movement of monarch butterfly larvae. *Environmental Entomology* 36: 228-233.

Prasifka JR, **Hellmich** RL, **Dively** GP, **Higgins** LS, **Dixon** PM & **Duan** JJ 2008. Selection of nontarget arthropod taxa for field research on transgenic insecticidal crops: using empirical data and statistical power. *Environmental Entomology* 37(1): 1-10.

Prihoda KR & **Coats** JR 2008. Aquatic fate and effects of *Bacillus thuringiensis* Cry3bb1 protein: Toward risk assessment. *Environmental Toxicology and Chemistry* 27(4): 793–798.

Ramirez-Romero R, **Chaufaux** J & **Pham-Delègue** MH 2005. Effects of Cry1Ab protoxin, deltamethrin and imidacloprid on the foraging activity and the learning performances of the honeybee *Apis mellifera*, a comparative approach. *Apidologie* 36: 601-611.

Ramirez-Romero R, **Bernal** JS, **Chaufaux** J & **Kaiser** L 2007. Impact assessment of Bt-maize on a moth parasitoid, *Cotesia marginiventris* (Hymenoptera: Braconidae), via host exposure to purified Cry1Ab toxin or Bt-plants. *Crop*

74

Protection 26: 953-962.

Ramirez-Romero R, **Desneux** N, **Decourtye** A, **Chaffiol** A & **Pham-Delègue** MH 2008. Does Cry1Ab toxin affect learning performances of the honey bee *Apis mellifera* L. (Hymenoptera, Apidae). *Ecotoxicology and Environmental Safety* 70: 327-333.

Raps A, **Kehr** J, **Gugerli** P, **Moar** WJ, **Bigler** F & **Hilbeck** A 2001. Immunological analysis of phloem sap of *Bacillus thuringiensis* corn and of the nontarget herbivore *Rhopalosiphum padi* (Homoptera: Aphididae) for the presence of Cry1Ab. *Molecular Ecology* 10: 525-533.

Rauschen S, **Schultheis** E, **Pagel-Wieder** S, **Schuphan** I & **Eber** S 2009. Impact of Bt-corn MON88017 in comparison to three conventional lines on *Trigonotylus caelestialium* (Kirkaldy) (Heteroptera: Miridae) field densities. *Transgenic Research* 18: 203-214.

Riddick EW, **Dively** G & **Barbosa** P 1998. Effect of a seed-mix deployment of Cry3A-transgenic and nontransgenic potato on the abundance of *Lebia grandis* (Coleoptera: Carabidae) and *Coleomegilla maculata* (Coleoptera: Coccinellidae). *Annals of the Entomological Society of America* 91(5): 647-653.

Rischer H & **Oksman-Caldentey** KM 2006. Unintended effects in genetically modified crops: revealed by metabolomics? *Trends in Biotechnology* 24(3): 102-104.

Rodrigo-Simón A, de Maagd RA, Avilla C, Bakker PL, Molthoff J, González-Zamora & Ferré J 2006. Lack of detrimental effects of *Bacillus thuringiensis* Cry toxins on the insect predator *Chrysoperla carnea*: a toxicological, histopathological, and biochemical analysis. *Applied and Environmental Microbiology* 72: 1595-1603.

Romeis J, **Dutton** A & **Bigler** F 2004. *Bacillus thuringiensis* toxin (Cry1Ab) has no direct effect on larvae of the green lacewind *Chrysoperla carnea*

(Stephens) (Neuroptera: Chrysopidae). *Journal of Insect Physiology* 50: 175-183.

Romeis J, **Meissle** M & **Bigler** F 2006. Transgenic crops expressing *Bacillus thuringiensis* toxins and biological control. *Nature Biotechnology* 24: 63-71.

Romeis J, Bartsch D, Bigler F, Candolfi MP, Gielkens MMC, Hartley SE, Hellmich RL, Huesing JE, Jepson PC, Layton R, Quemada H, Raybould A, Rose RI, Schiemann J, Sears MK, Shelton AM, Sweet J, Vaituzis Z, & Wolt JD 2008. Assessment of risk of insect-resistant transgenic crops to non-target arthropods. *Nature Biotechnology* 26: 203-208.

Romeis J, **Meissle** M, **Raybould** A & **Hellmich** RL 2009. Environmental impact of genetically modified crops on above-ground non target arthropods. In: *Environmental Impact of Genetically Modified Crops*, Ferry N & Gatehouse AMR (eds.). CAB International, UK. pp. 165-198.

Rose R, **Dively** GP & **Pettis** J 2007. Effects of Bt corn pollen on honey bees: Emphasis on protocol development. *Apidologie* 38: 368-377.

Rosi-Marshall EJ, **Tank** JL, **Royer** TV, **Whiles** MR, **Evans-White** M, **Chambers** C, **Griffiths** NA, **Pokelsek** J & **Stephen** ML 2007. Toxins in transgenic crop by products may affect headwater stream ecosystems. *Proceedings of the National Academy of Sciences USA* 104: 16204-16208.

Saxena D & **Stotzky** G 2001a. *Bacillus thuringiensis* (Bt) toxin released from root exudates and biomass of Bt corn has no apparent effect on earthworms, nematodes, protozoa, bacteria, and fungi in soil. *Soil Biology* & *Biochemisty* 33: 1225-1230.

Saxena D & **Stotzky** G 2001b. Bt corn has a higher lignin content than non-Bt corn. *American Journal of Botany* 88(9): 1704–1706.

Saxena D, Flores S & Stotzky G 2002. Bt toxin is released in root exudates

from 12 transgenic corn hybrids representing three transformation events. *Soil Biology & Biochemistry* 34: 133-137.

Schaefer CW & **Panizzi** AR 2000. *Heteroptera of economic importance*. CRC Press, Boca Raton, Florida, USA.

Schmidt JEU, Braun CU, Whitehouse LP & Hilbeck A 2009. Effects of activated Bt transgene products (Cry1Ab, Cry3Bb) on immature stages of the ladybird Adalia bipunctata in laboratory ecotoxicity testing. Archives of Environmental Contamination and Toxicology 56: 221-228.

Schrader S, **Münchenberg** T, **Baumgarte** S & **Tebbe** CC 2008. Earthworms of different functional groups affect the fate of the Bt-toxin Cry1Ab from transgenic maize in soil. *European Journal of Soil Biology* 44: 283-289.

Schuler TH, Potting RPJ, Denholm I, Clark SJ, Clark AJ, Stewart CN, & Poppy GM. (2003). Tritrophic choice experiments with Bt-plants, the diamondback moth (*Plutella xylostella*) and the parasitoid *Cotesia plutellae*. *Transgenic Research* 12:351-361.

Sears MK, Hellmich RL, Siegfried BD, Pleasants JM, Stanly-Horn DE, Oberhauser KS & Dively GP (2001). Impact of Bt corn pollen on monarch butterfly populations: a risk assessment. *Proceedings of the National Academy of Sciences USA* 98: 11937-11942.

Secretariat of the Convention on Biological Diversity 1992. Convention on Biological Diversity. Montreal, Canada. Available at http://www.biodiv.org/doc/legal/cbd-en.pdf. Accessed online March 2010.

Sims SR & **Martin** JW 1997. Effect of *Bacillus thuringiensis* insecticidal proteins Cry1A(b), Cry1A(c), CryIIA, and CryIIIA on *Folsomia candida* and *Xenylla grisea* (Insecta: Collembola). *Pedobiologia* 41: 412-416.

Symondson WOC, **Sunderland** KD & **Greenstone** MH 2002. Can generalist predators be effective biocontrol agents? *Annual Review of Entomology* 47: 561–594.

Tarkalson DD, **Kachman** SD, **Knops** JMN, **Thies** JE & **Wortmann** CS 2008. Decomposition of Bt and non-Bt corn hybrid residues in the field. *Nutrient Cycling in Agroecosystems* 80: 211-222.

Todd JH, **Ramankutty** P, **Barraclough** EI & **Malone** LA 2008. A screening method for prioritizing non-target invertebrates for improved biosafety testing of transgenic crops. *Environmental Biosafety Research* 7: 35–56.

Torres JB & **Ruberson** JR 2008. Interactions of *Bacillus thuringiensis* Cry1Ac toxin in genetically engineered cotton with predatory heteropterans. *Transgenic Research* 17: 345-354.

Turrini A, Sbrana C, Nuti MP, Pietrangeli BM & Giovanetti M 2004. Development of a model system to assess the impact of genetically modified corn and aubergine plants on arbuscular mycorrhizal fungi. *Plant Soil* 266: 69-75.

USA EPA 1998. Guidelines for Ecological Risk Assessment. United States Environmental Protection Agency (USA EPA). *Federal Register* 63(93): 26846–26924. Available at http://cfpub.epa.gov/ncea/cfm/recordisplay. cfm?deid=12460. Accessed online March 2010.

Van Toan P, Hoàng NB, Anyango B, Zwahlen C, Manachini B, Andow DA, Wheatley RE 2008. Potential effects of Transgenic cotton on Soil Ecosystem Processes in Vietnam. In: Environmental Risk Assessment of Genetically Modified Organisms: Vol. 4. Challenges and opportunities with Bt Cotton in Vietnam. Andow D, Hilbeck A & Van Tuàt N (eds.). CABI Publishing, Wallingford, UK. pp 258-273.

Vercesi ML, **Krogh** PH & **Holmstrup** M 2006. Can *Bacillus thuringiensis* (Bt) corn residues and Bt-corn plants affect life-history traits in the earthworm *Aporrectodea caliginosa? Applied Soil Ecology* 32: 180-187.

Verkerk. KJ 2004. Manipulation of tritrophic interactions in IPM. In: Integrated Pest Management: Potential, constraints and challenges. Koul O, Dhaliwal GS & Cuperus GW (eds.). CABI Publishing, UK. pp 55-72.

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Wandeler H, **Bahylova** J & **Nentwig** W 2002. Consumption of two Bt and six non-Bt corn varieties by the woodlouse *Porcellio scaber*. *Basic and Applied Ecology* 3: 357-365.

Weber M & **Nentwig** W 2006. Impact of Bt corn on the diplopod *Allajulus latestriatus*. *Pedobiologia* 50: 357-368.

Widmer F 2007. Assessing effects of transgenic crops on soil microbial communities. Advances in Biochemical Engineering and Biotechnology 107: 207-234.

Wold SJ, **Burkness** EC, **Hutchinson** WD & **Venette** RC 2001. In-field monitoring of beneficial insect populations in transgenic corn expressing a *Bacillus thuringiensis* toxin. *Journal of Entomological Science* 36(2): 177-187.

Wolfenbarger LL, **Naranjo** SE, **Lundgren** JG, **Bitzer** RJ & **Watrud** LS 2008. Bt crop effecs on functional guilds of non-target arthropods: a meta-analysis. *PLoS ONE* 3: 1-11 (e2118).

Wraight SP & **Ramos** ME 2005. Synergistic interaction between *Beauveria* bassiana- and *Bacillus thuringiensis tenebrionis*-based biopesticides applied against field populations of Colorado potato beetle larvae. *Journal* of Invertebrate Pathology 90: 139–150.

Xue K, **Luo** HF, **Qi** HY & **Zhang** HX 2005. Changes in soil microbial community structure associated with two types of genetically engineered plants analyzing by PLFA. *Journal of Environmental Science (China)* 17: 130–134.

Yao HW, Ye GY, Jiang CY, Fan LJ, Datta K, Hu C & Datta SK 2006. Effect of the pollen of transgenic rice line TT9-3 with a fused cry1Ab/cry1Ac gene from *Bacillus thuringiensis* Berliner on non-target domestic silkworm *Bombix mori* Linnaeus (Lepidoptera: Bombyxidae). *Applied Entomology* and *Zoology* 41: 339-348.

Zhang GF, Wan FH, Liu WX & Guo JY 2006a. Early instar response to plant-

delivered Bt-toxin in a herbivore (*Spodoptera litura*) and a predator (*Propylaea japonica*). Crop Protection 25: 527–533.

Zhang GF, **Wan** FH, **Lövei** GA, **Liu** WX & **Guo** JY 2006b. Transmission of Bt toxin to the predator *Propylaea japonica* (Coleoptera: Coccinellidae) through its aphid prey feeding on transgenic Bt cotton. *Environmental Entomology* 35(1): 143-150.

Zwahlen C, **Hilbeck** A, **Howald** R & **Nentwig** W 2003. Effects of transgenic Bt corn litter on earthworm *Lumbricus terrestris. Molecular Ecology* 12: 1077-1086.

Zwahlen C, **Hilbeck** A, **Nentwig** W 2007. Field decomposition of transgenic Bt-maize residue and the impact on non-target soil invertebrates. *Plant Soil* 300: 245-257.